



MITOCHONDRIAL BIOGENESIS AND REDOX REGULATION

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MITOCHONDRIAL BIOGENESIS AND REDOX REGULATION

○ Objectives

- Provide an overview of the physiological and pathological states and transcriptional control mechanisms involved in mitochondrial biogenesis
- Define how specific adjustments in cellular and/or mitochondrial redox state activate mitochondrial biogenesis
- Illustrate the role of redox-regulation of mitochondrial biogenesis in the resolution of mitochondrial damage during acute inflammation



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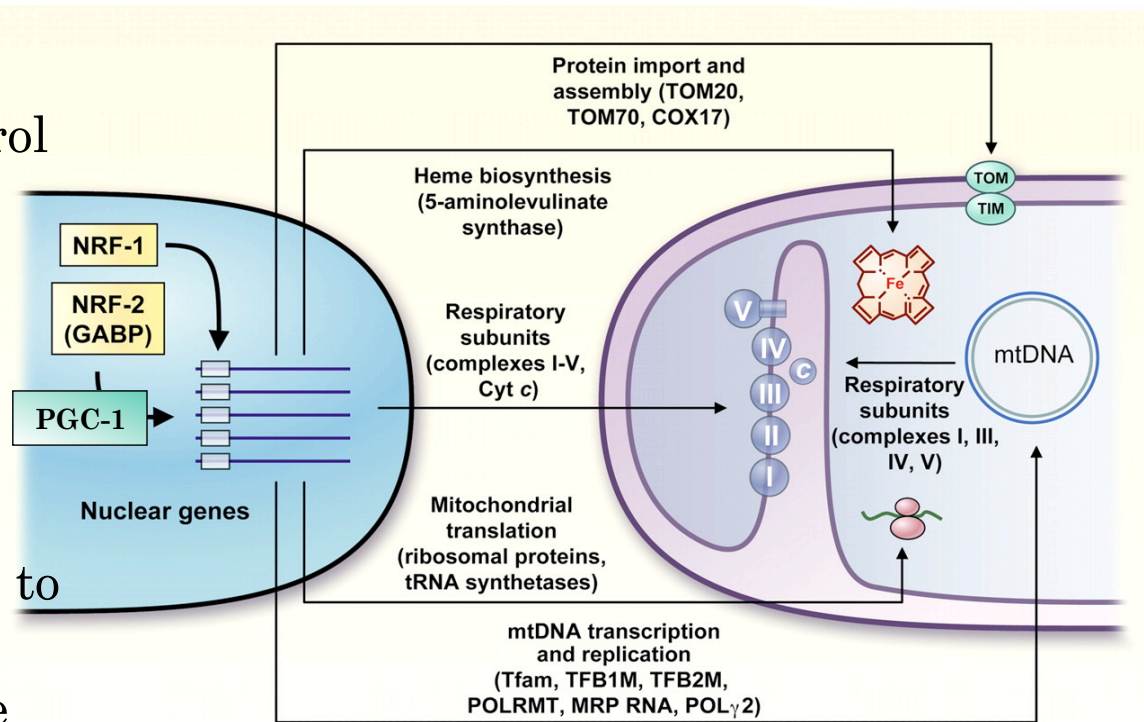
○ List of Abbreviations

- AMPK: AMP-activated kinase
- CREB: Cyclic AMP-responsive element-binding protein 1
- ERRα: Estrogen-related receptor alpha
- Keap1: Kelch-like ECH-associated protein 1
- NAMPT: nicotinamide phosphoribosyltransferase
- Nfe2l2: Nuclear factor erythroid 2-related factor 2 (Nrf2)
- NRF-1/NRF-2: Nuclear respiratory factors-1 and -2 (GABP)
- PGAM5: Mitochondrial serine/threonine phosphatase
- PGC-1α: Peroxisome proliferator-activated receptor gamma co-activator 1-alpha
- PPARα: Peroxisome proliferator-activated receptor alpha
- RIP140: Nuclear receptor-interacting protein 1
- SIRT1: NAD-dependent deacetylase sirtuin-1



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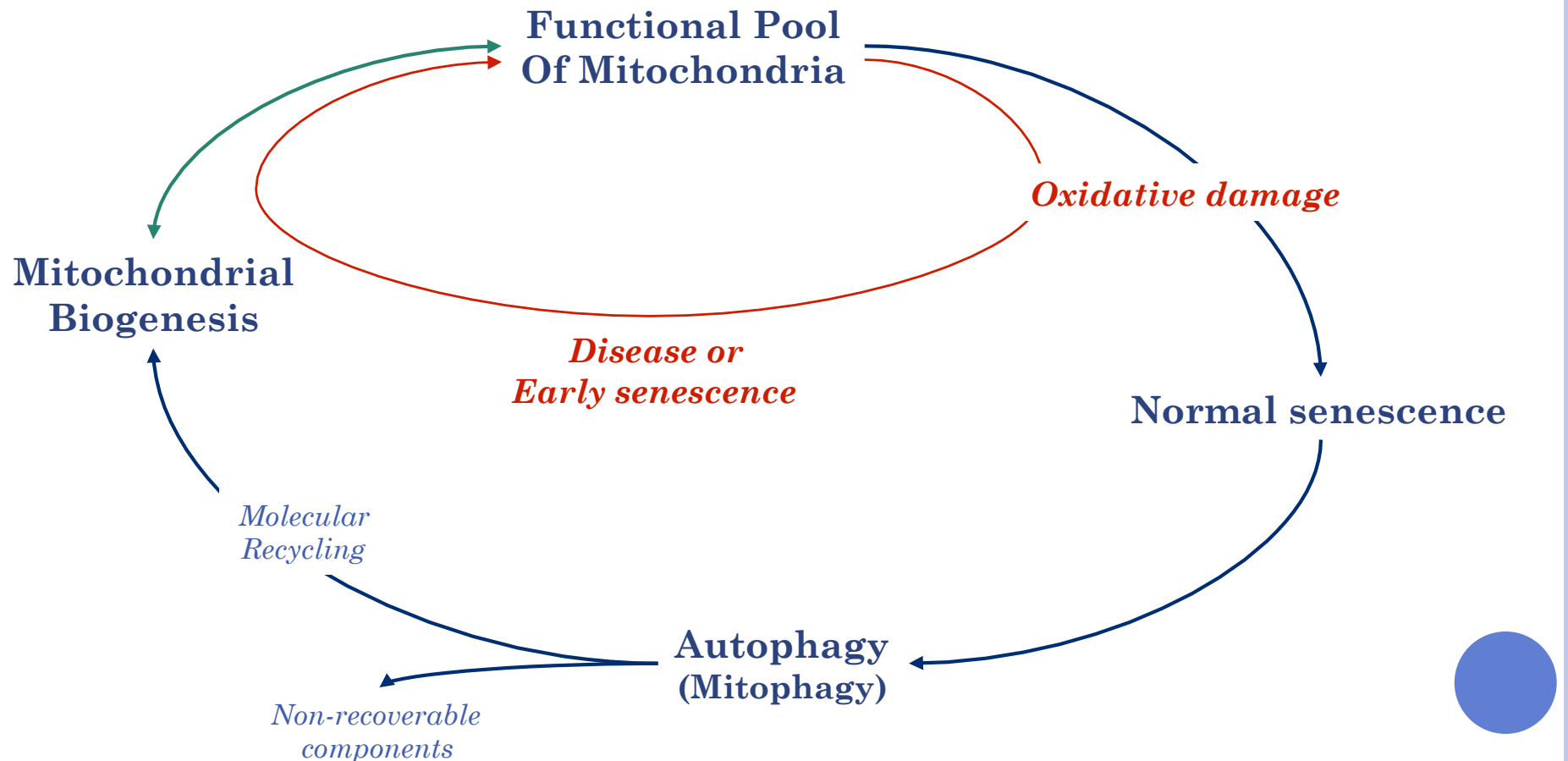
- Bi-genomic transcriptional network integral to mitochondrial quality control
- Coordinated with energy needs, cell growth, proliferation, autophagy
- Homeostatic, adaptive, anti-oxidant, anti-inflammatory
- Part of integrated response to tissue damage and disease
- Major control pathways are under redox-regulation



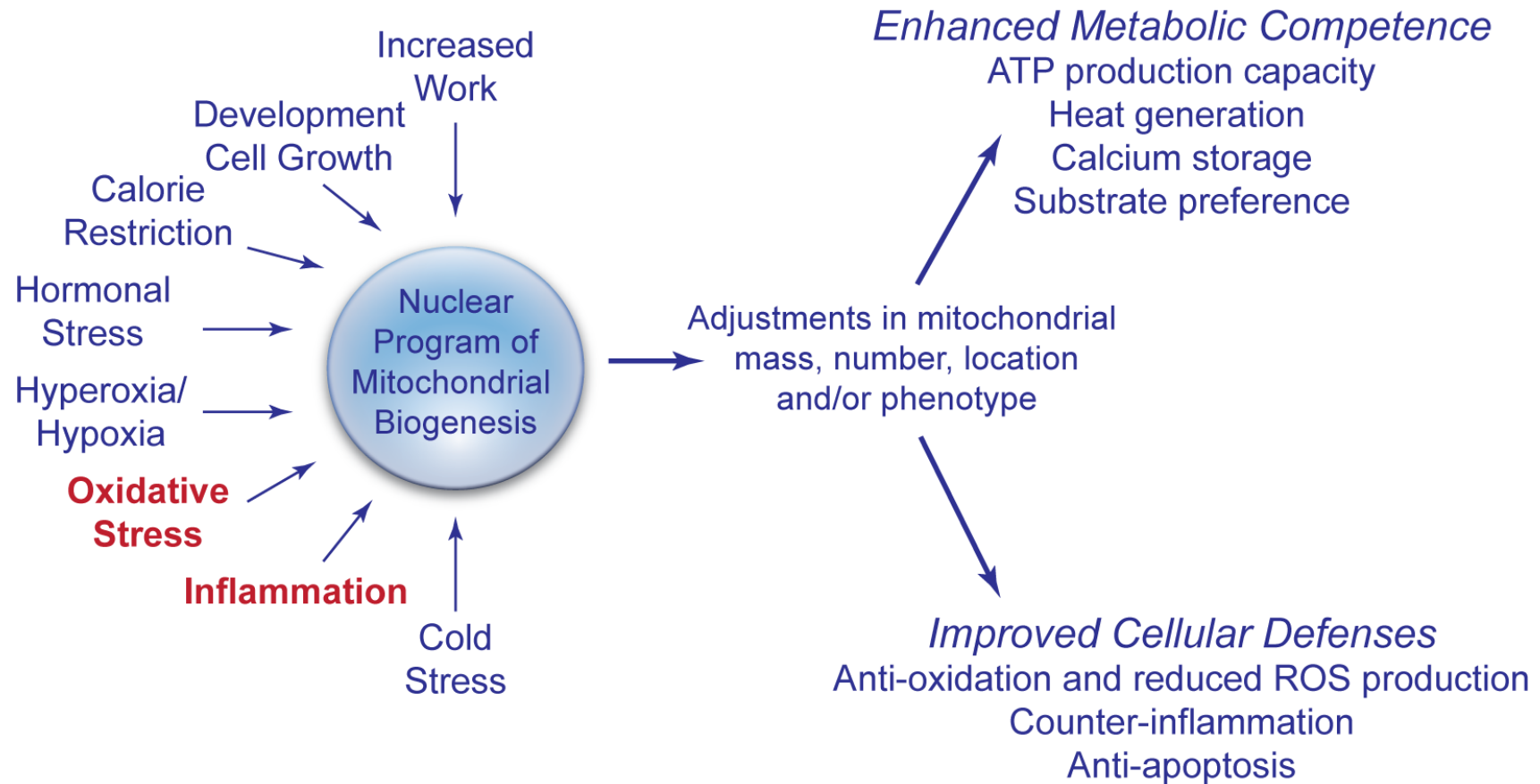
Scarpulla, R. *Physiol Rev* 2008;88:611-638

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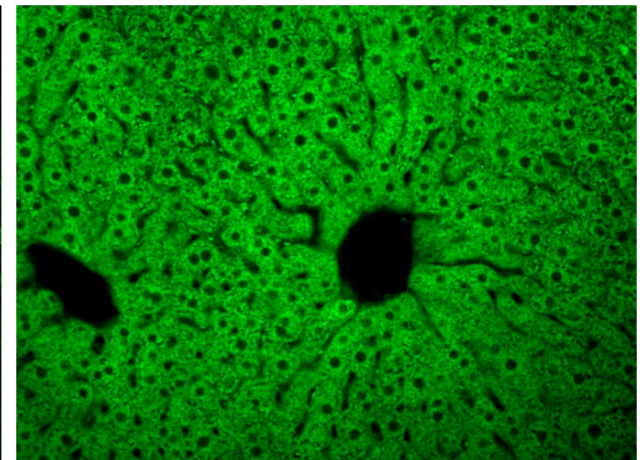
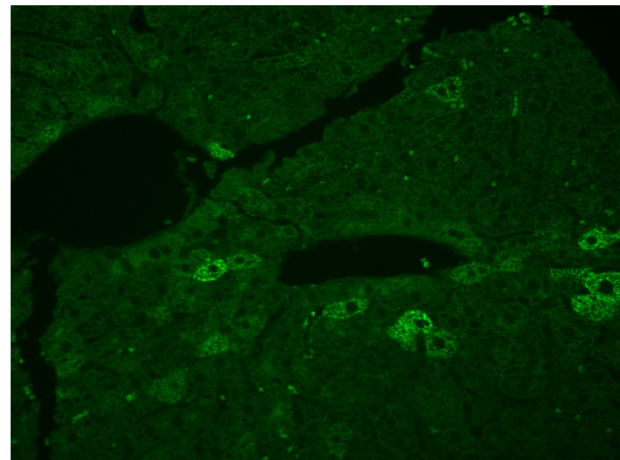
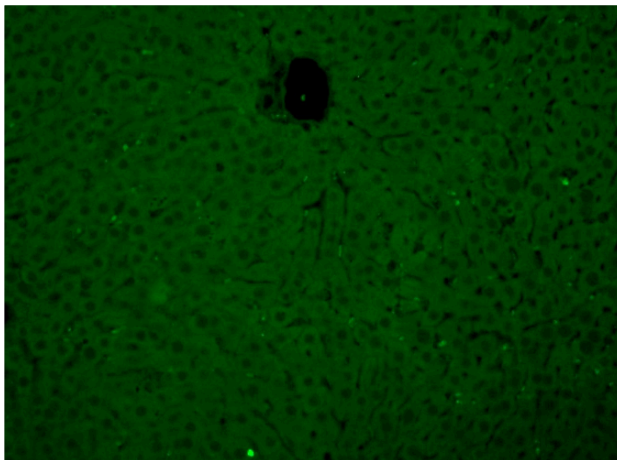
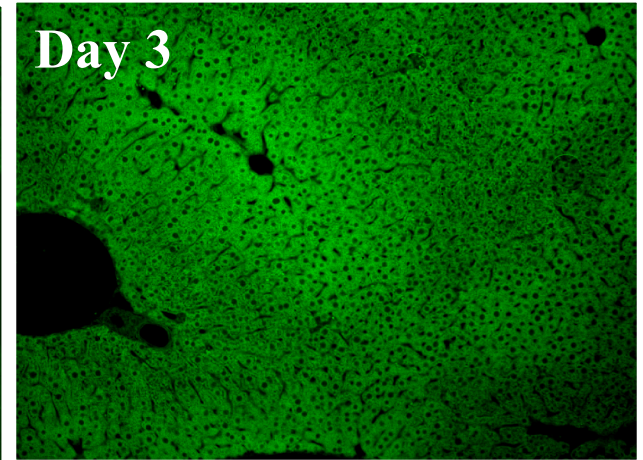
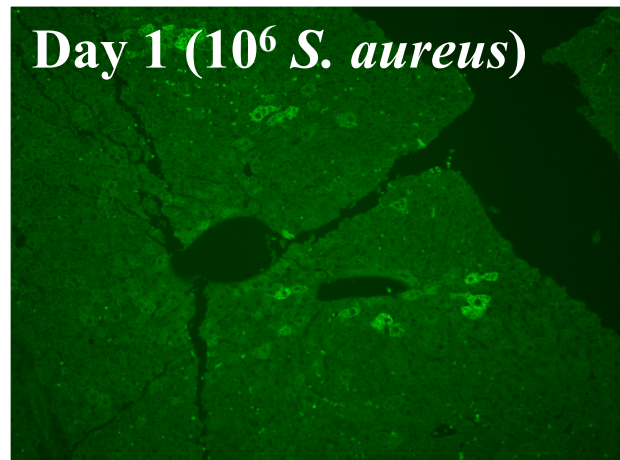
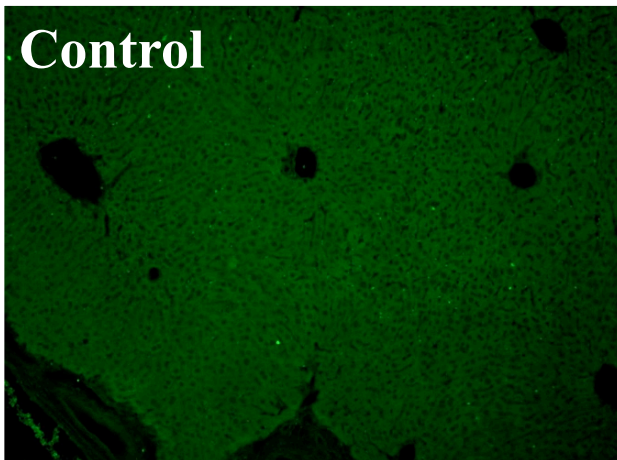
- Mitochondrial Quality Control (no *de novo* synthesis)



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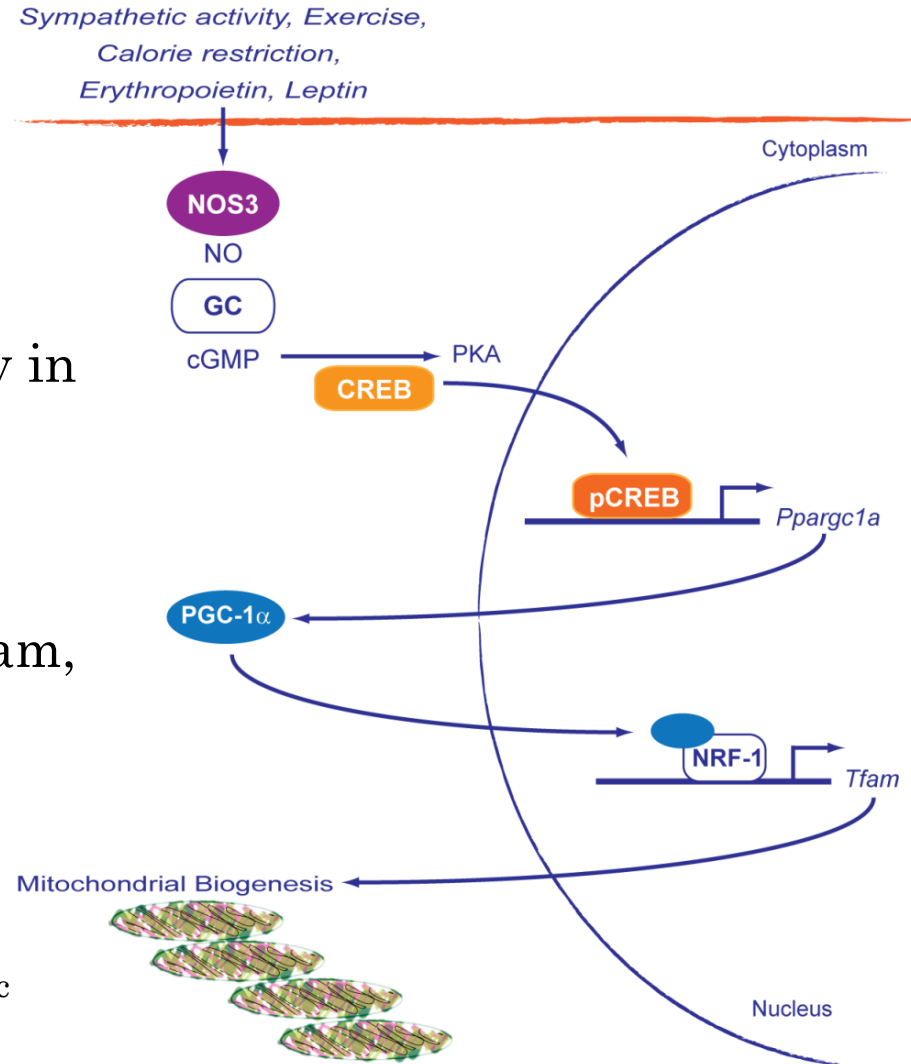
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- Major types of induction mechanisms for nuclear-encoded mitochondrial genes
 - Energy-sensing pathways
 - AMPK
 - **SIRT1**
 - **CREB**
- Ca⁺⁺-dependent kinases
 - CaMK II/IV, CaMKK
 - Calcineurin A
 - **p38g MAPK**
- Redox-regulated pathways
 - **NO/cGMP**
 - **Nrf2/HO-1/CO**
- Inflammatory pathways
 - **NF-kB**
 - **CREB**



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- NO and mitochondrial biogenesis
 - cGMP production by multiple factors either directly or through an increase in eNOS activity in muscle, fat, other tissues up-regulates regulatory genes for mitochondrial biogenesis including PGC-1 α , NRF-1, and Tfam, leading to mitochondrial proliferation



Modified from Nisoli E, Carruba MO. Nitric oxide and mitochondrial biogenesis. *J Cell Sci.* 2006;119:2855–2862.

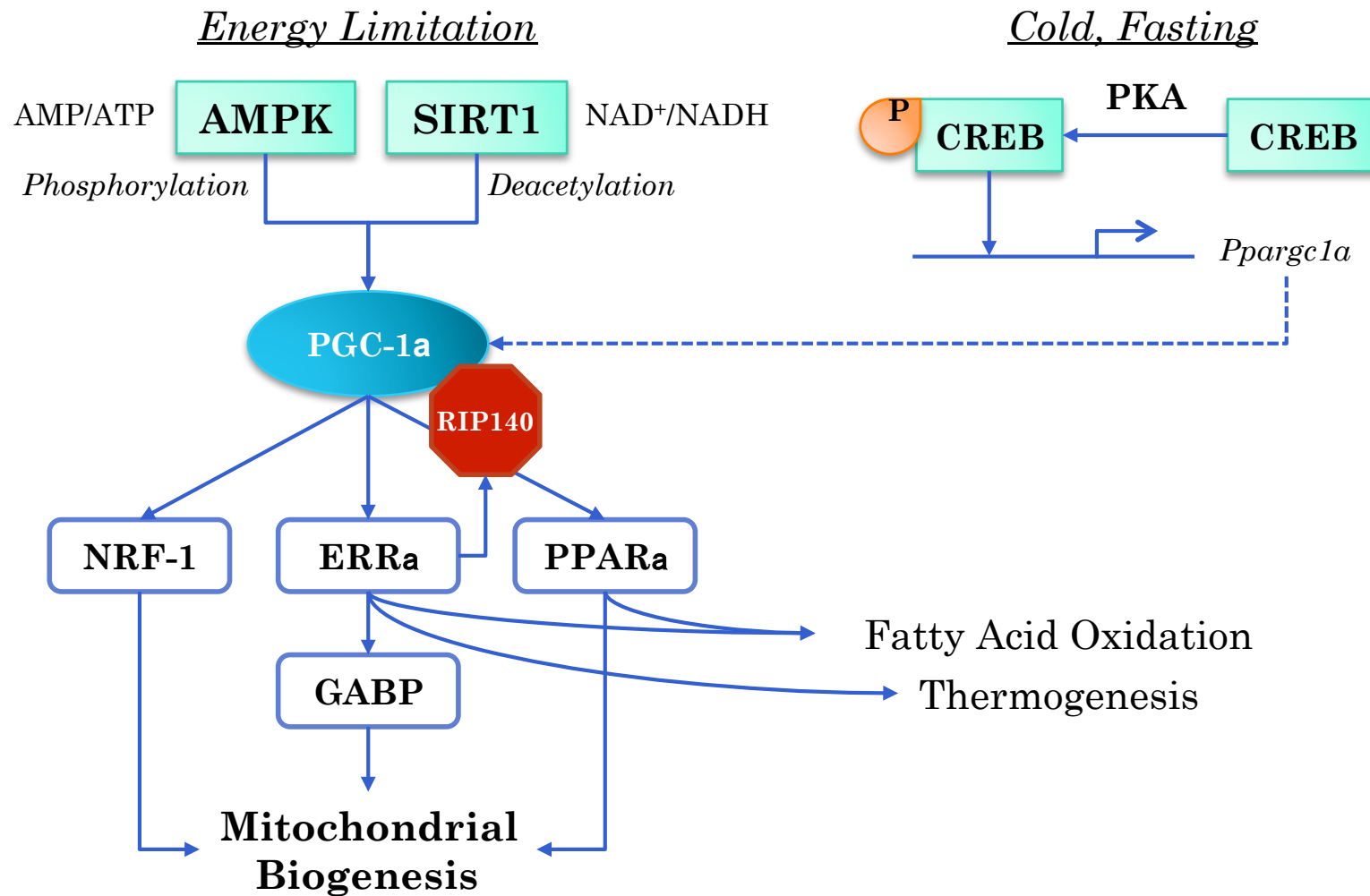
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- PGC-1 co-activator family members act as coordinators of mitochondrial biogenesis
 - PGC-1a
 - PGC-1b
 - PRC
- Partner with DNA binding transcription factors (e.g. NRF-1, GABP and CREB) to activate 500-1,000 nuclear-encoded mitochondrial genes needed for mitochondrial proliferation

Vercauteren K., Pasko R.A., Gleyzer N., Marino V.M., Scarpulla R.C. PGC-1-related coactivator: immediate early expression and characterization of a CREB/NRF-1 binding domain associated with cytochrome c promoter occupancy and respiratory growth. *Mol. Cell. Biol.* 26:7409-7419, 2006



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○ SIRT1

- NAD⁺-dependent histone deacetylase (class III HDAC)
- Deacetylates diverse histone and non-histone proteins
- Mediates effect of calorie restriction on lifespan and cell metabolism
- Anti-inflammatory effect through down-regulation of NF-κB-dependent pro-inflammatory cytokine expression



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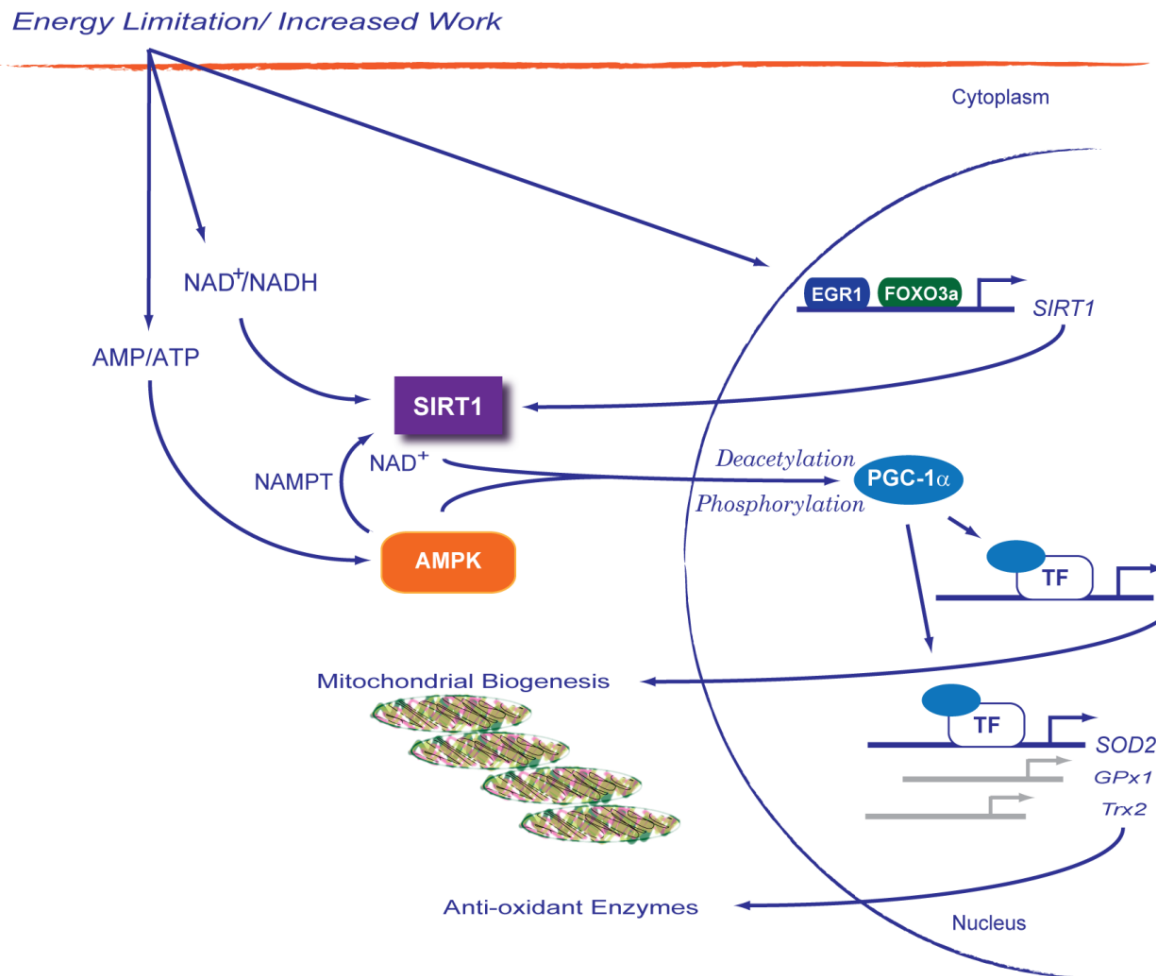
○ AMPK

- Senses metabolic stress and energy deprivation (AMP/ATP); activated by muscle contraction (Ca^{2+} /ROS)
- Stimulates fatty acid breakdown, glycolysis
- Inhibits energy-utilizing synthetic pathways (protein, cholesterol, fatty acids)
- Activates PGC-1 α by phosphorylating Thr¹⁷⁷ and Ser⁵³⁸
- Activates SIRT1 by increasing NAD^+/NADH through β - oxidation and stimulation of nicotinamide phosphoribosyltransferase (NAMPT), the rate-limiting step in NAD biosynthesis



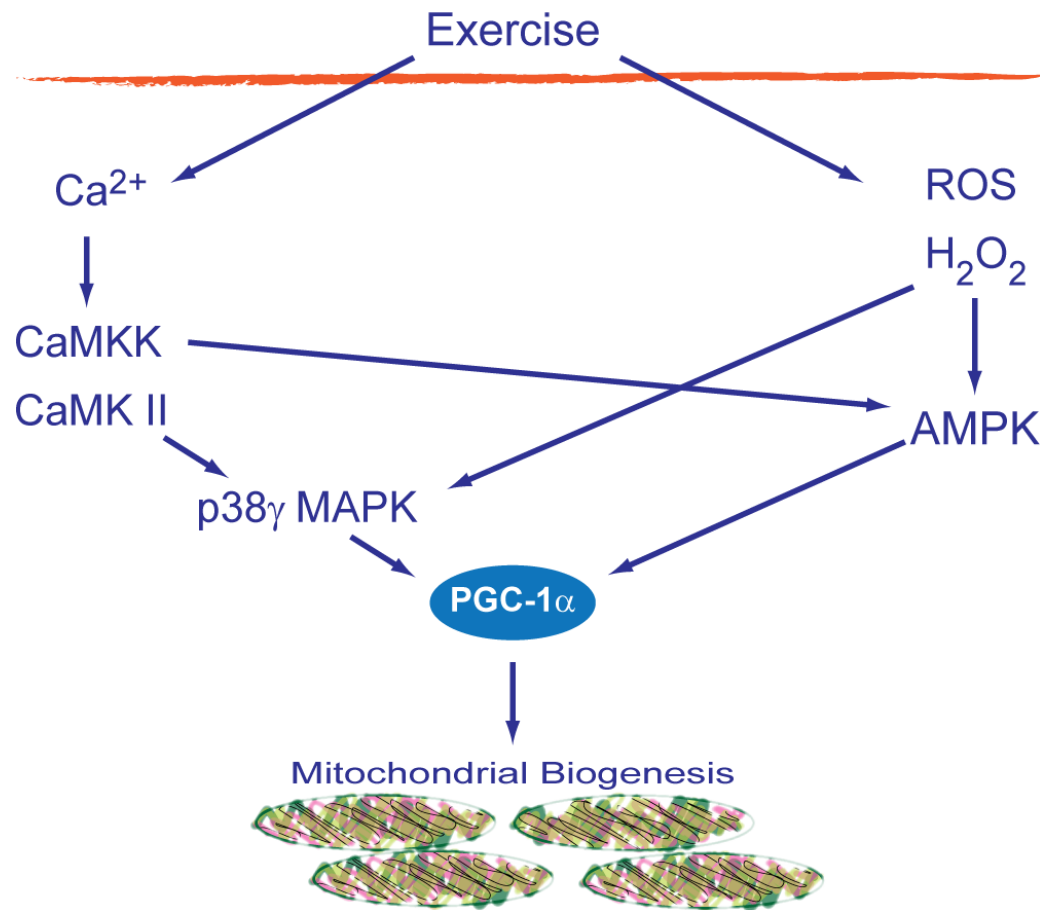
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- SIRT1 and AMPK cooperate to activate PGC-1α



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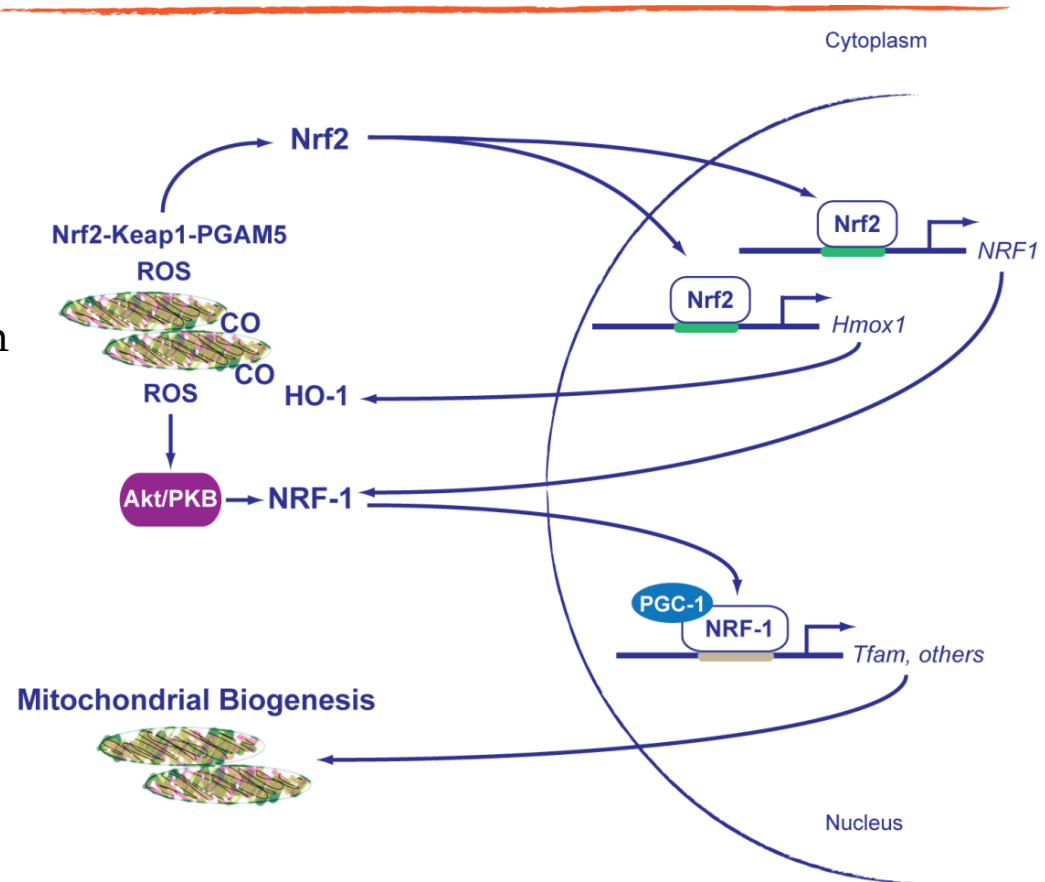
- Ca^{2+} vs. ROS in exercise-induced mitochondrial biogenesis



Modified from Lira V A *et al. Am J Physiol Endocrinol Metab* 299:E145-E161, 2010

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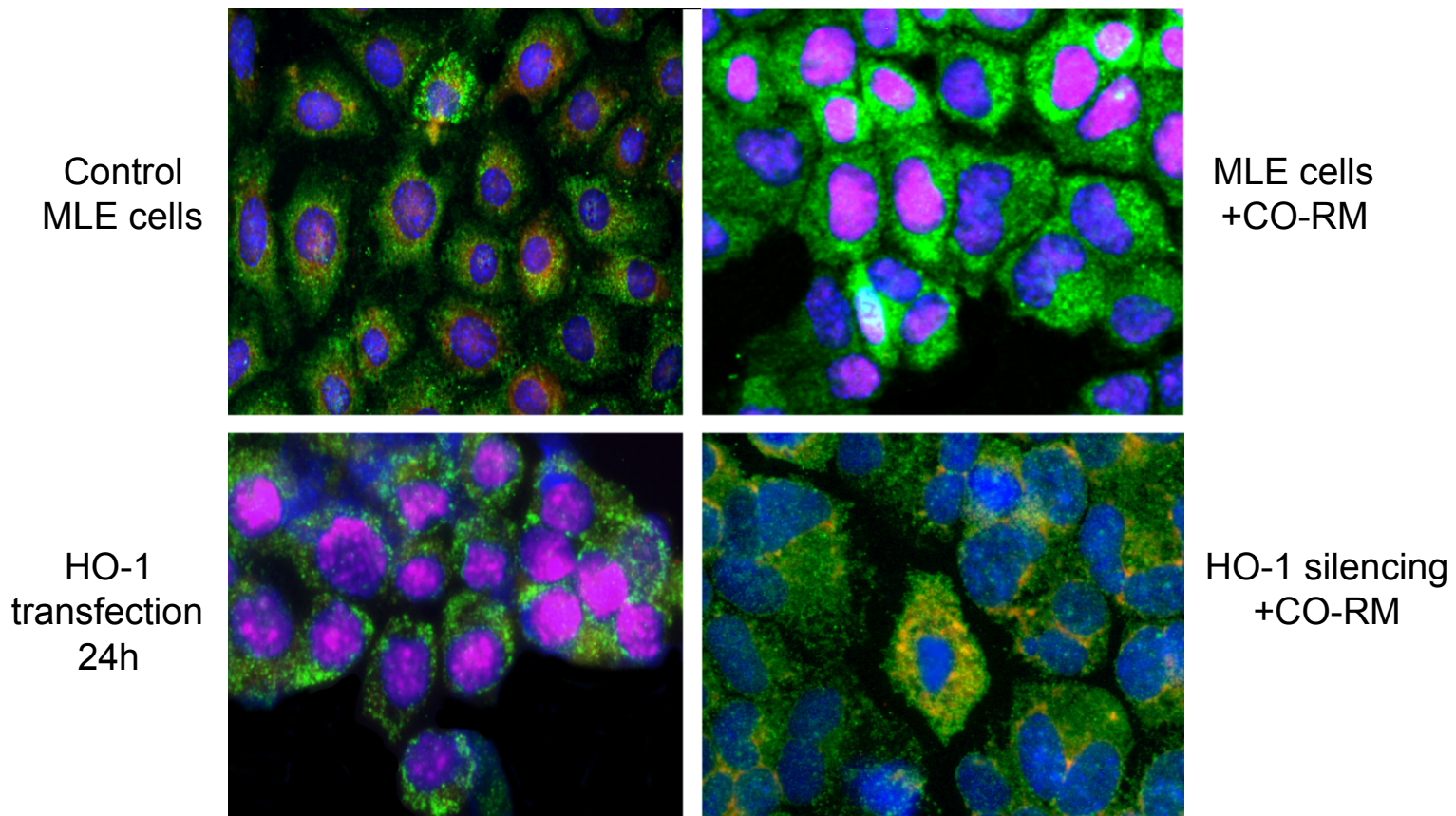
- Heme oxygenase-1 (HO-1; *Hmox1*)
 - Inducible heme catabolism
 - Breaks α -methene carbon bond releasing Fe, CO, biliverdin
 - Potent anti-inflammatory, pro-survival effects
 - HO-1/CO induces mitochondrial biogenesis



Suliman HB, *et al.* A new activating role for CO in cardiac mitochondrial biogenesis. *J Cell Sci.* 120:299-308, 2007

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- HO-1/CO induction of mitochondrial biogenesis



Green, Mitotracker; Red, NRF-1; Blue, Dapi

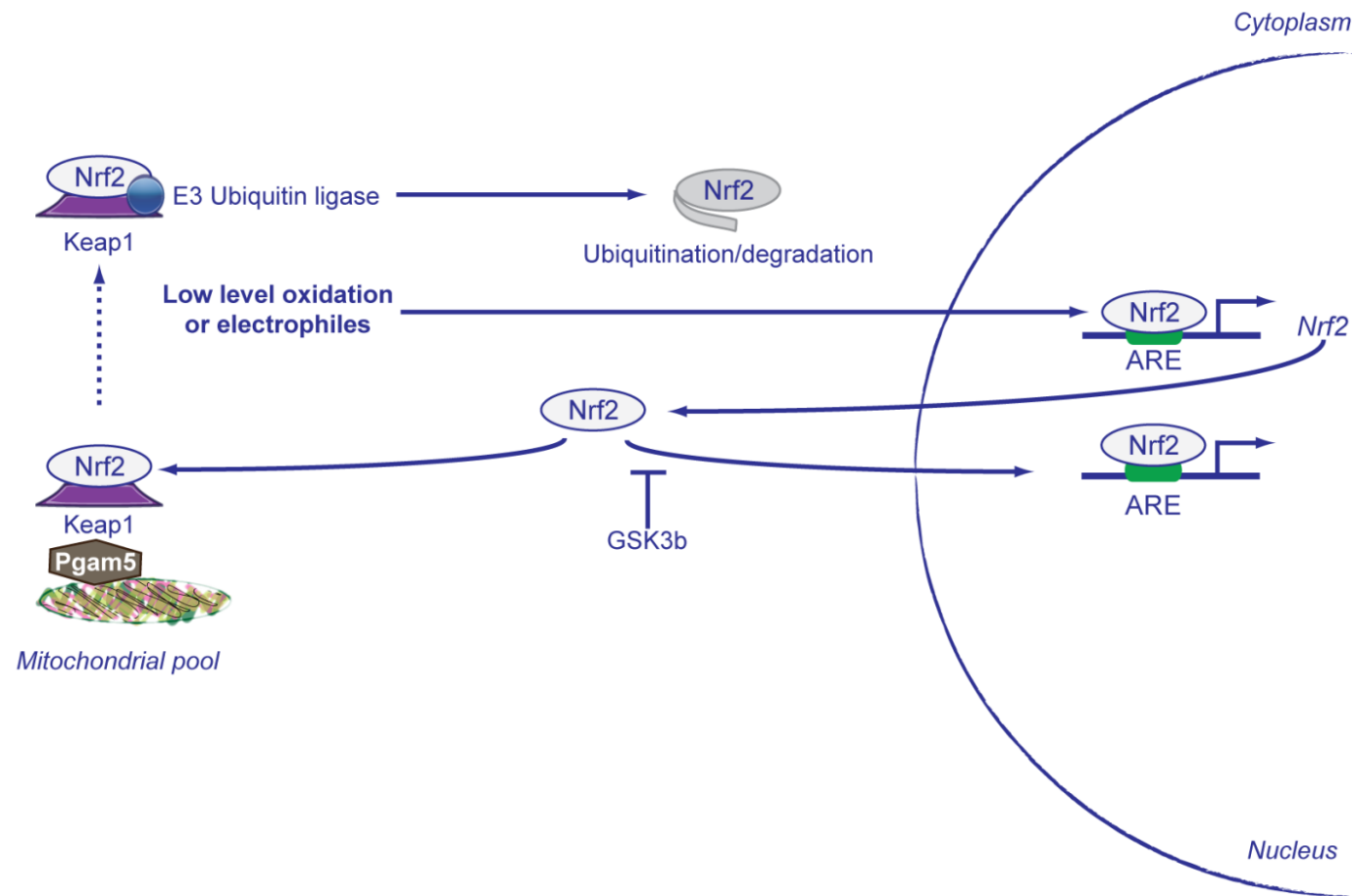
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- Key role of Nrf2 (Nfe2l2)
 - Basic leucine-zipper “CNC” transcription factor that binds to antioxidant response elements (ARE) in promoter regions of target genes, e.g. *Nqo1*, *GST*, *Hmox1*
 - Forms ternary docking complex with Pgam5 (outer mitochondrial membrane) and Keap1 (cytoplasmic inhibitor)
 - Keap1 cysteine oxidation stabilizes complex and prevents degradation
 - Nrf2 Ser⁴⁰ phosphorylation by PKC dissociates Nrf2 from Keap1 in response to oxidative stress
 - Widely expressed in kidney, muscle, lung, heart, liver, brain

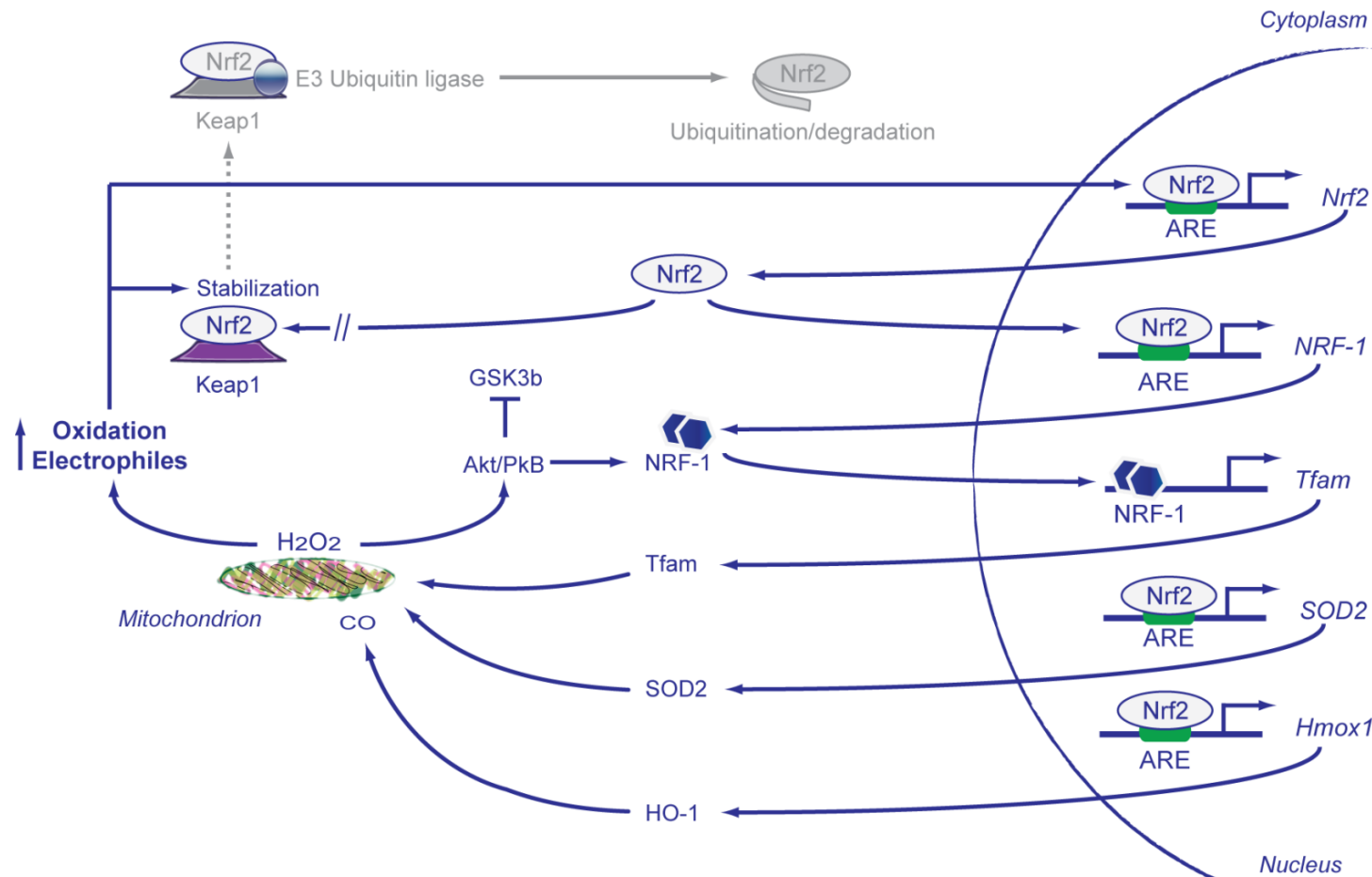


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- Normal cytoplasmic turnover of Nrf2



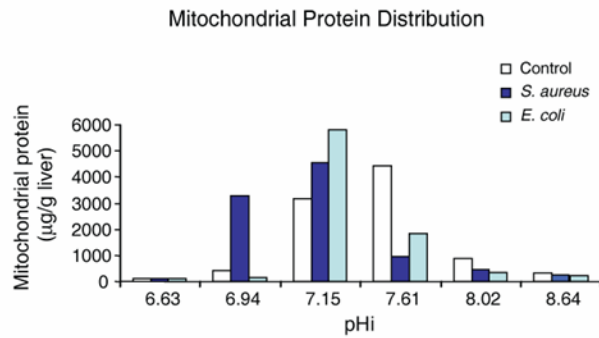
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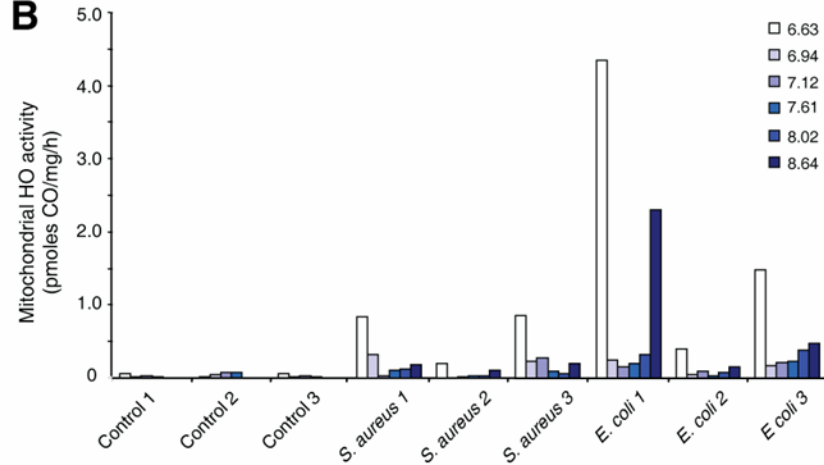
Piantadosi CA, *et al.* Heme oxygenase-1 regulates cardiac mitochondrial biogenesis via Nrf2-mediated transcriptional control of nuclear respiratory factor-1. *Circ Res.* 103(11):1232-40, 2008

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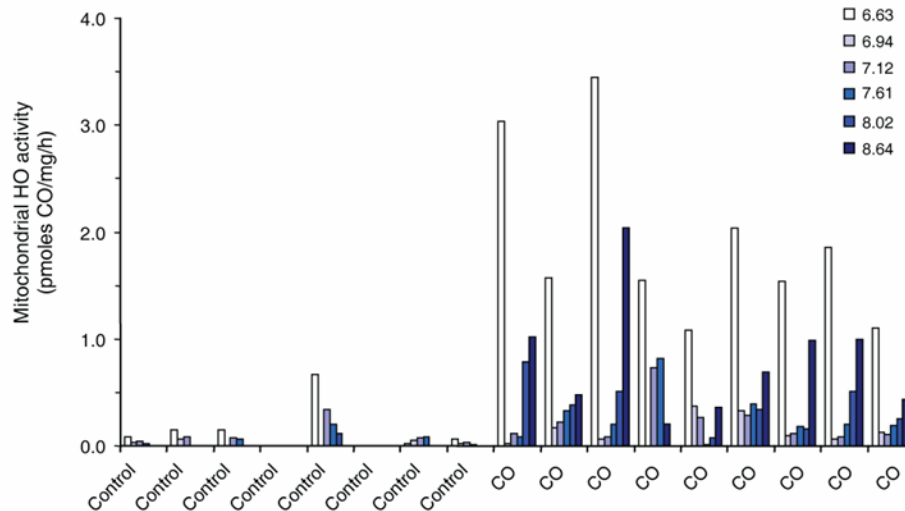
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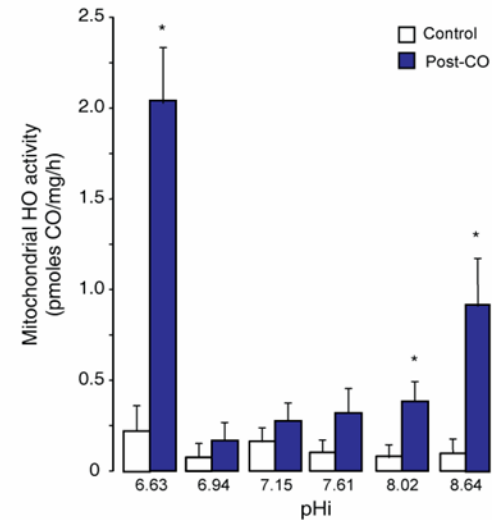
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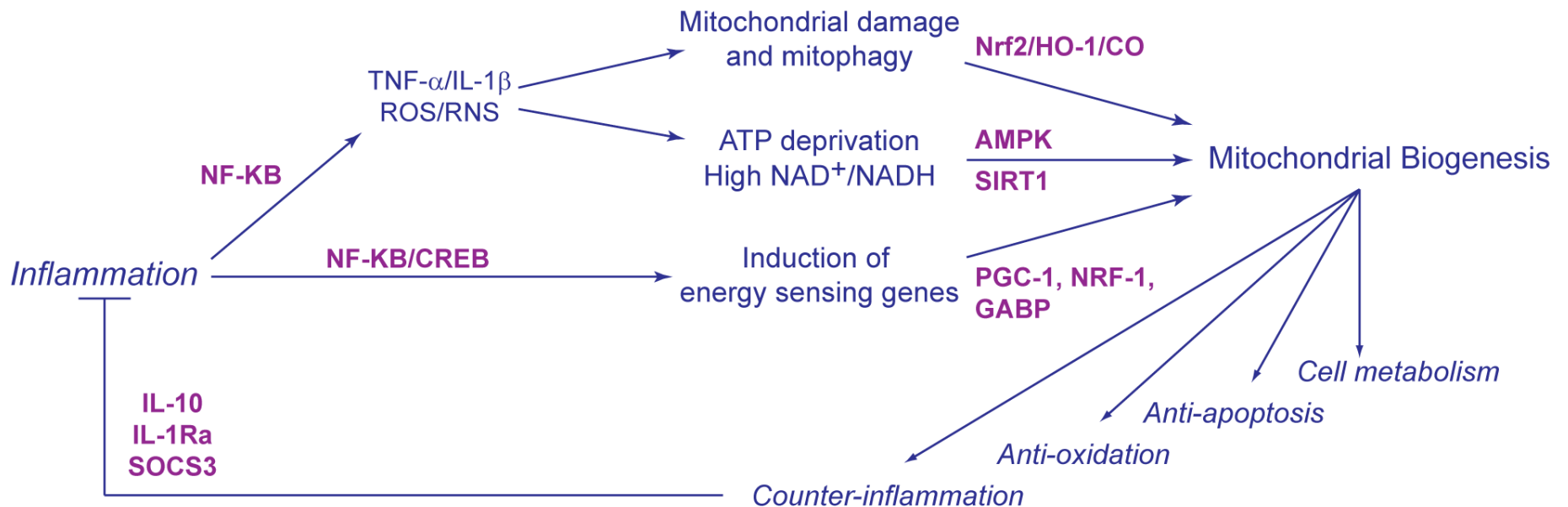


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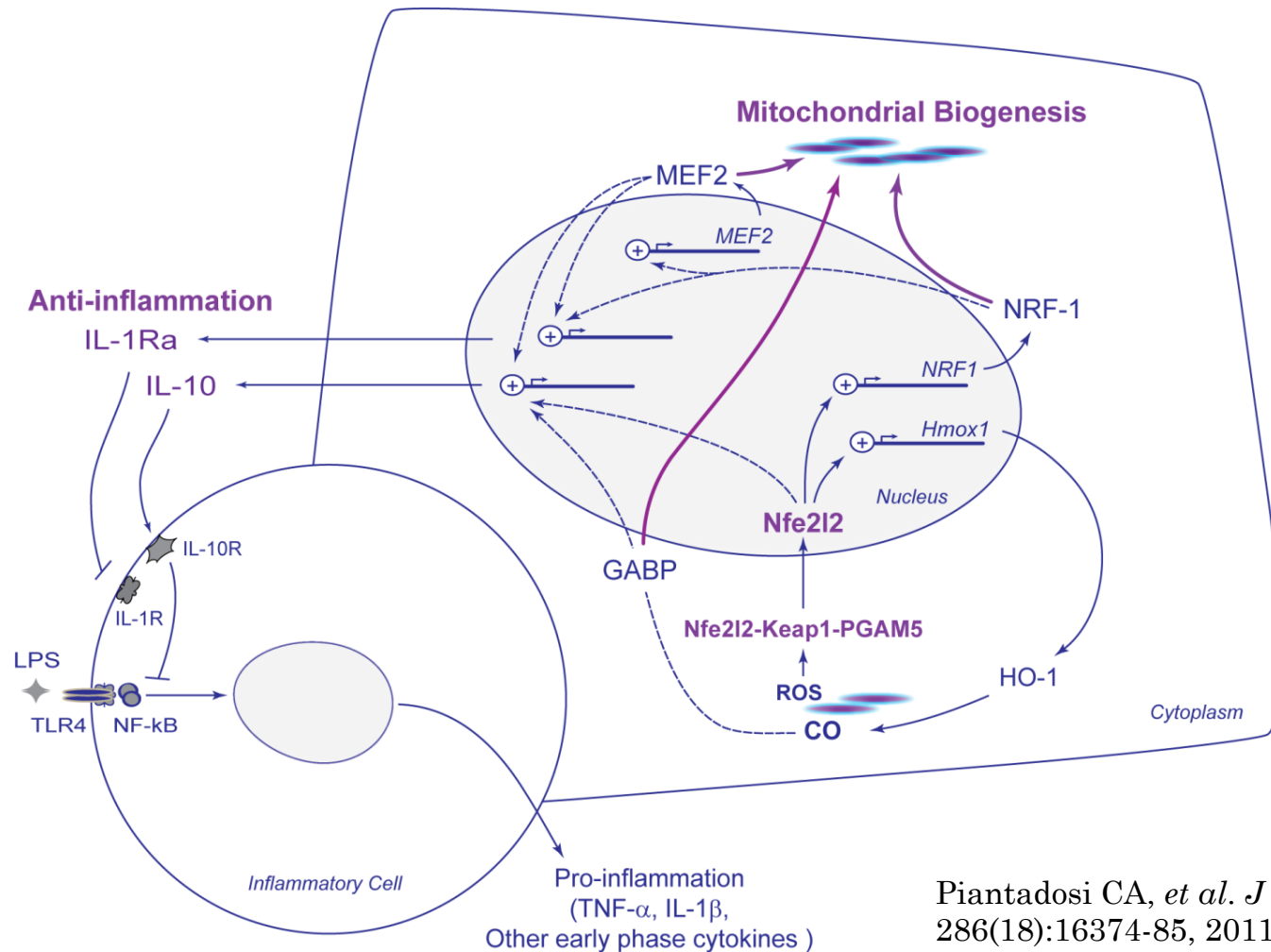
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- Effects of inflammation on mitochondrial quality control



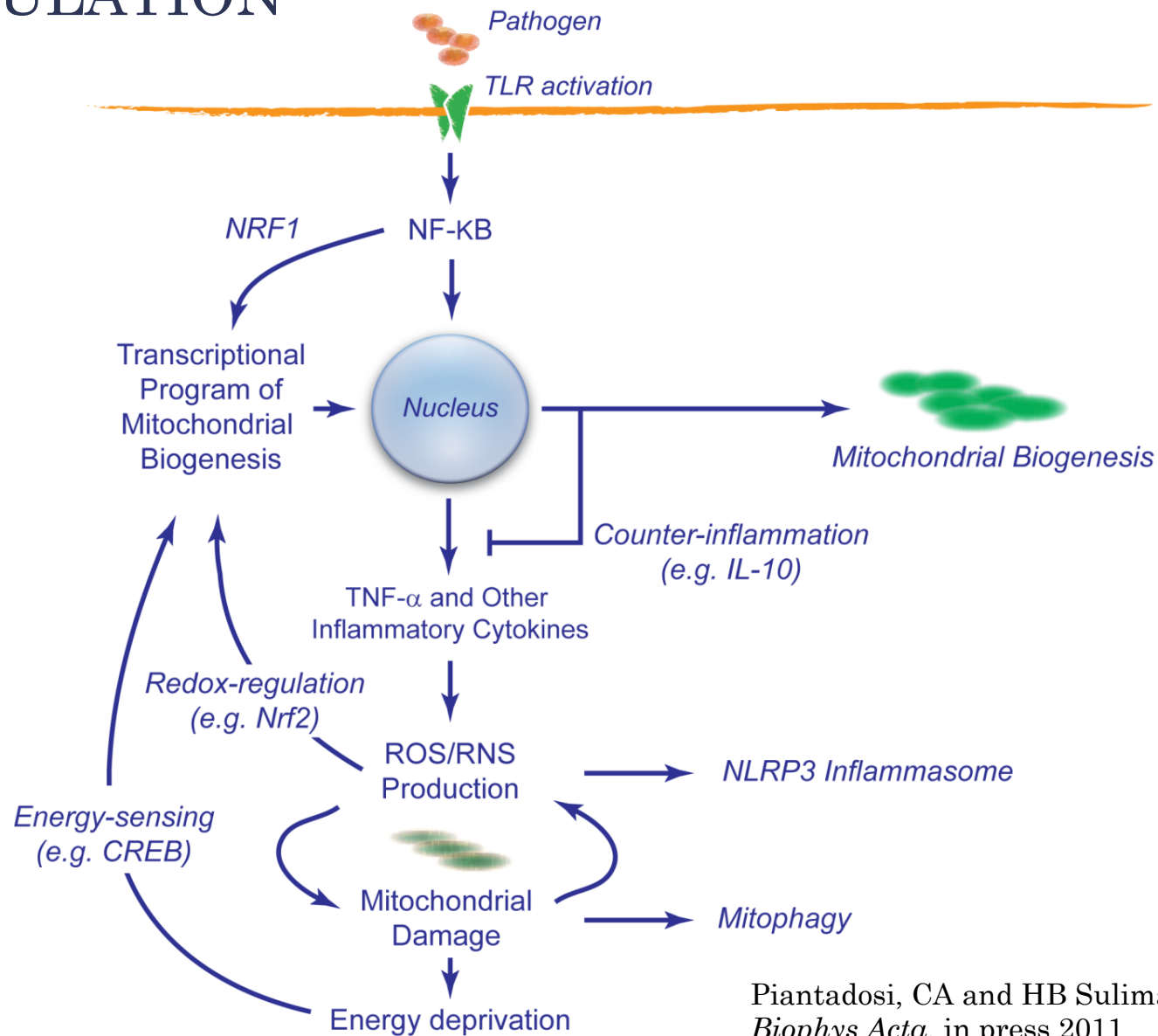
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○ Counter-inflammation



Piantadosi CA, et al. *J Biol Chem.* 286(18):16374-85, 2011

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Piantadosi, CA and HB Suliman, *Biochem Biophys Acta*, in press 2011

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○ Summary/Conclusions

- Redox regulation of mitochondrial biogenesis is a major mitochondrial quality control mechanism in mammalian cells
- Multiple levels of redox control are involved
 - NO/cGMP
 - Nrf2/HO-1/CO
 - NAD⁺/NADH ratio
 - Mitochondrial H₂O₂ production
- Control points involve redox regulation of kinase activity and gene expression
- The mitochondrial biogenesis transcriptional program is part of a master network of cellular anti-oxidant and anti-inflammatory defenses



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○ Collaborators

- Hagir B. Suliman, D.V.M, Ph.D.
- Karen E. Welty Wolf, M.D.
- Raquel B. Bartz, M.D.
- Timothy E. Sweeney, M.D., Ph.D.
- Crystal Withers, M.D., Ph.D. student
- Ping Fu, M.D.

○ Funding

- NHLBI
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