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# Physiological Fluxes of Reactive Oxygen Species During Inflammation: *Reality Check*

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# Physiological Fluxes of Reactive Oxygen Species During Inflammation: *Reality Check*



***Inflammation*** is a **protective**  
response designed to destroy  
invading pathogens

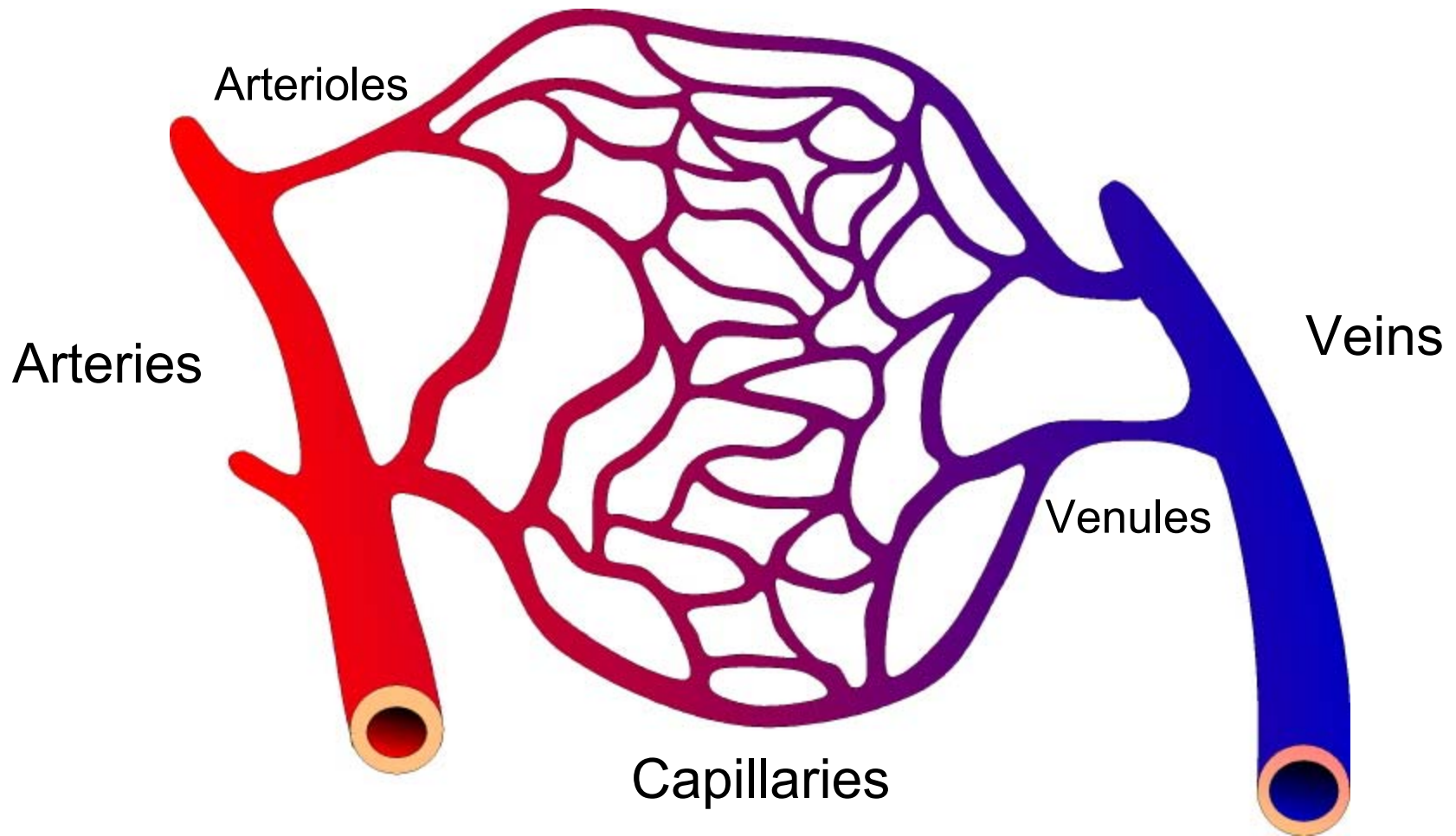
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- **Redness (rubor)**
- **Heat (calor)**
- **Swelling (tumor)**
- **Pain (dolor)**

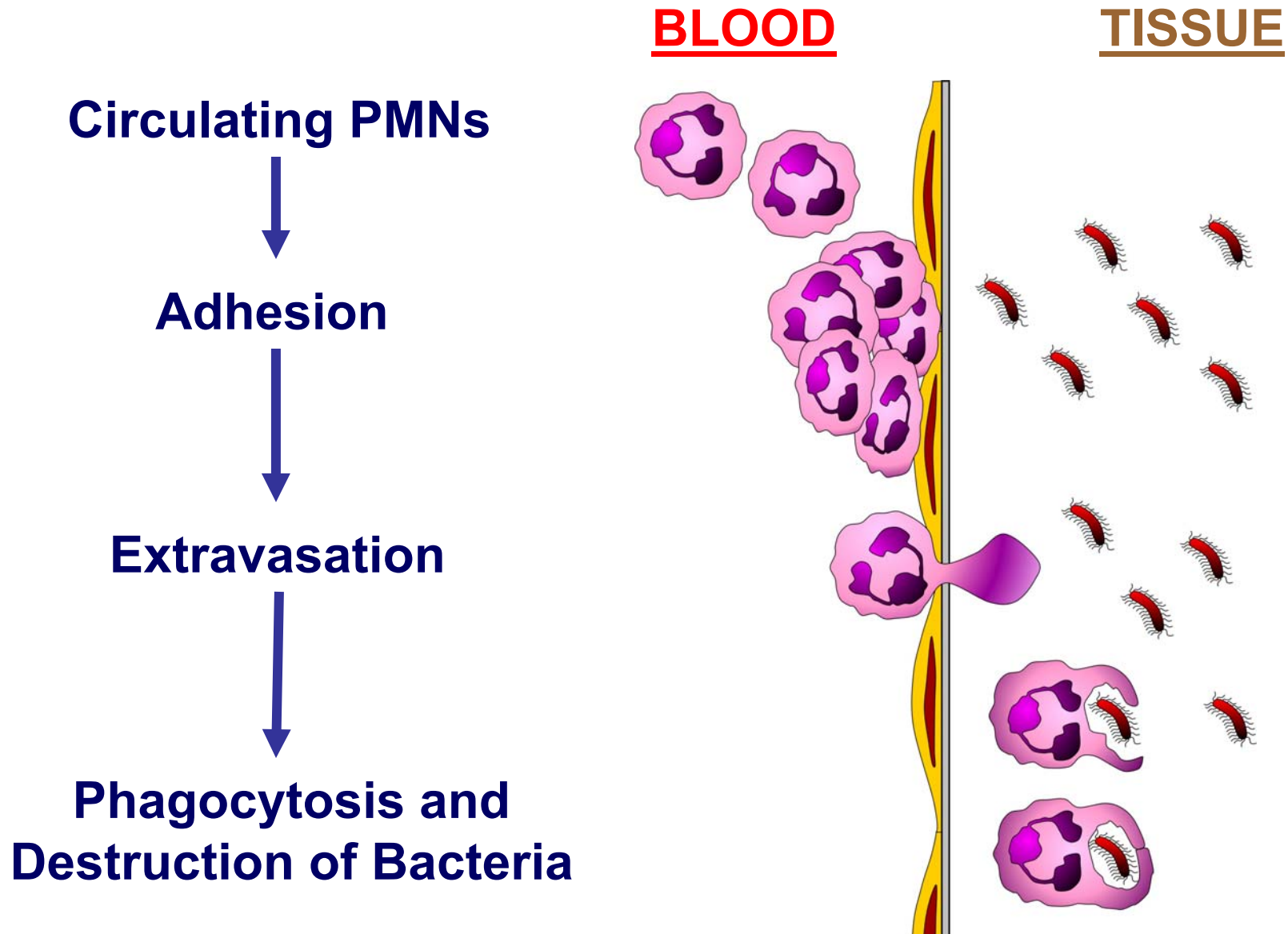
# ***Inflammation* is a Double-Edge Sword**

- ***Defects*** in mounting an inflammatory response may lead to grave illness or death.
- ***Excessive inflammation*** leads to tissue injury, pain and loss of function.

***Location:***  
**Tissue Microcirculation**



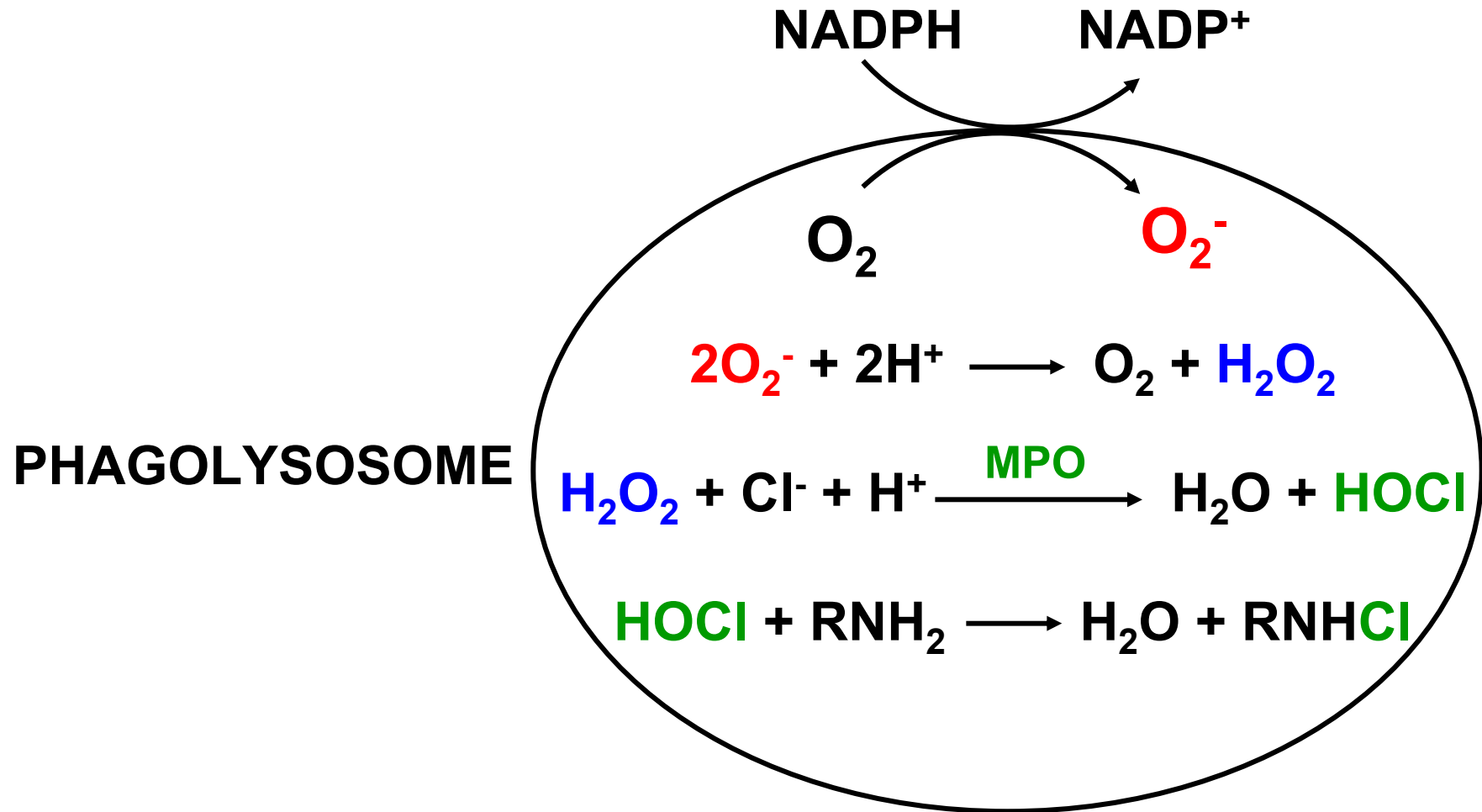
# PMN-Endothelial Cell Interactions In Response to Infection: Post-Capillary Venules





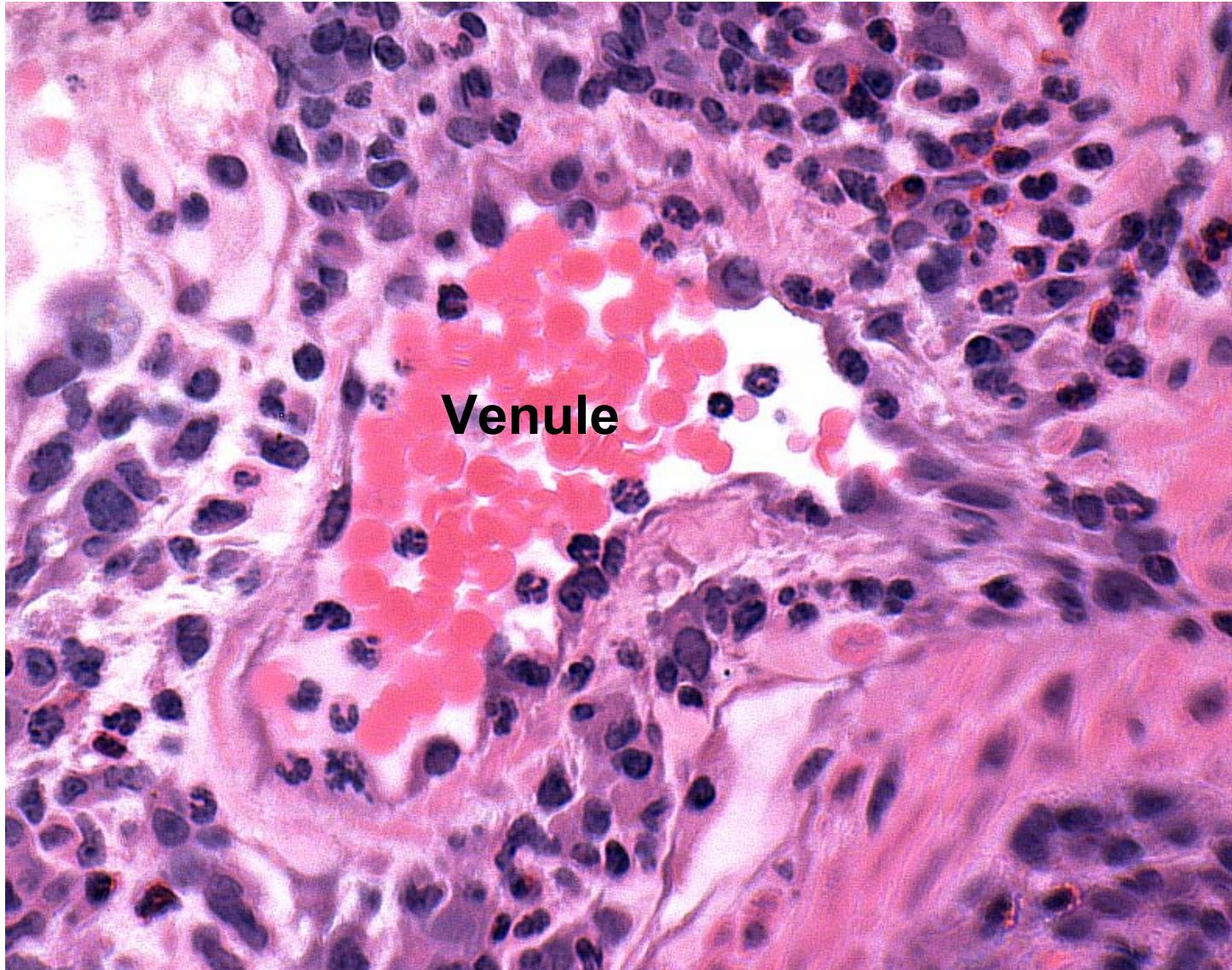


# OXIDATIVE BURST



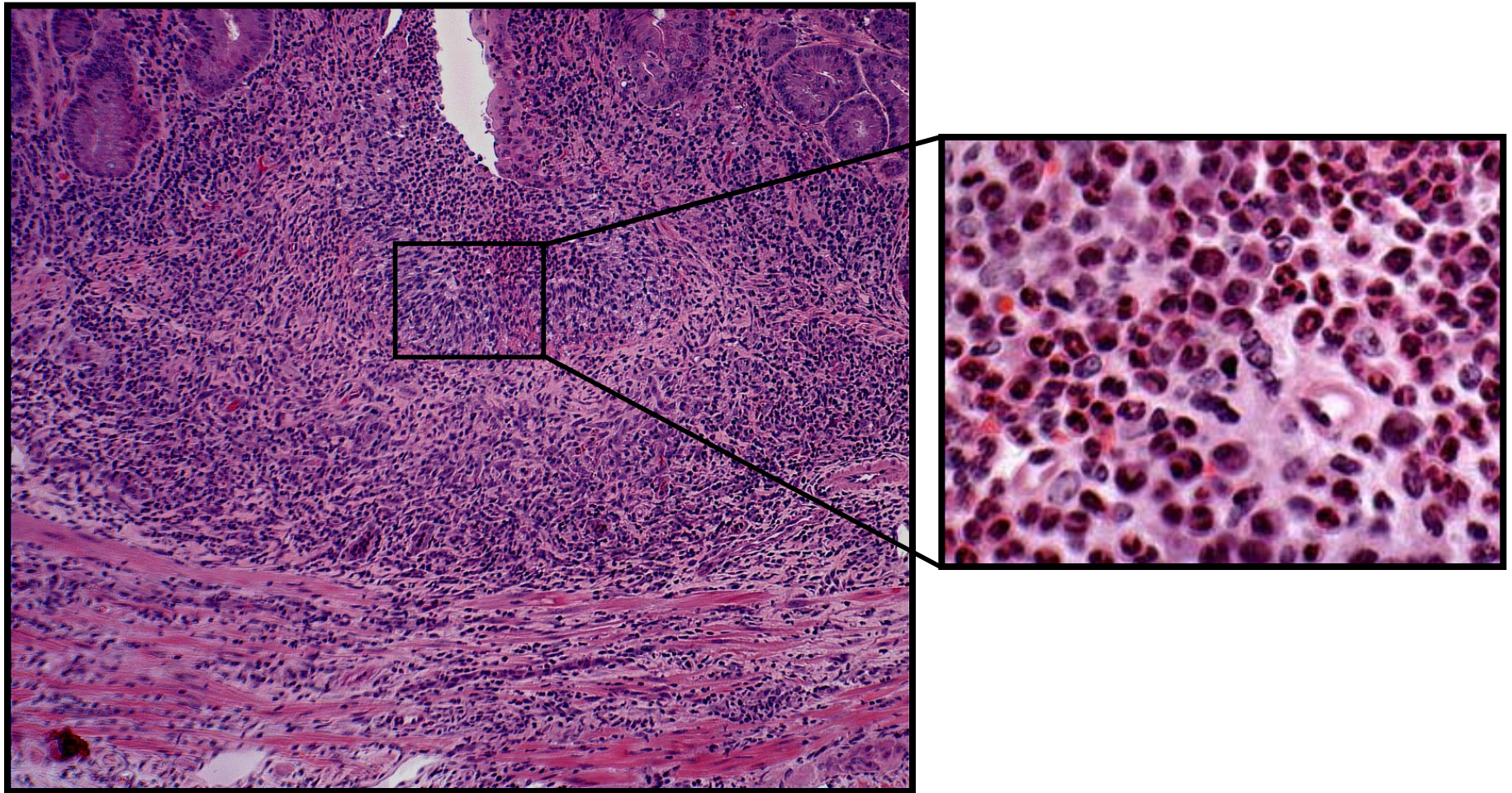


# Large Numbers of PMNs Invade Tissue During an Inflammatory Response

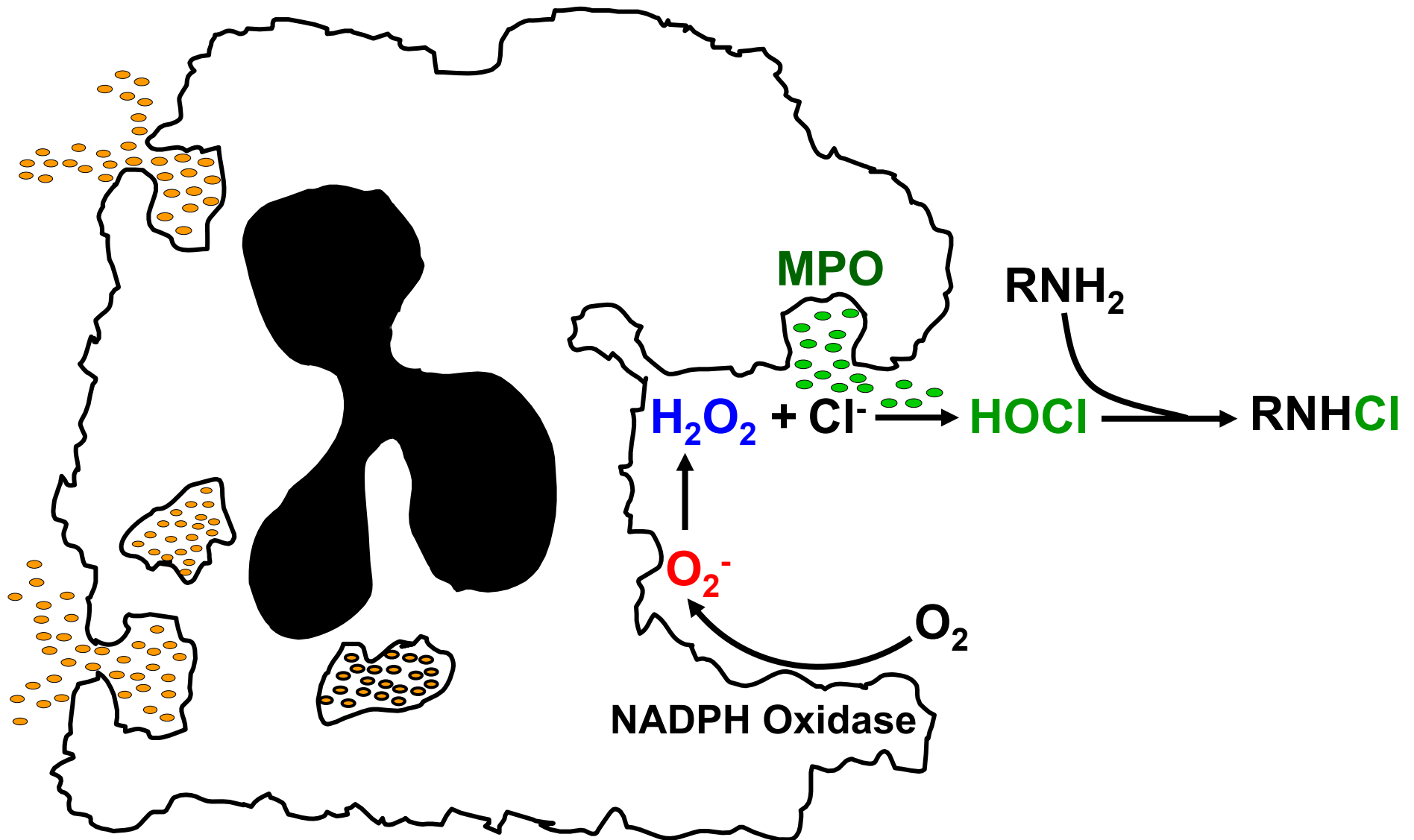




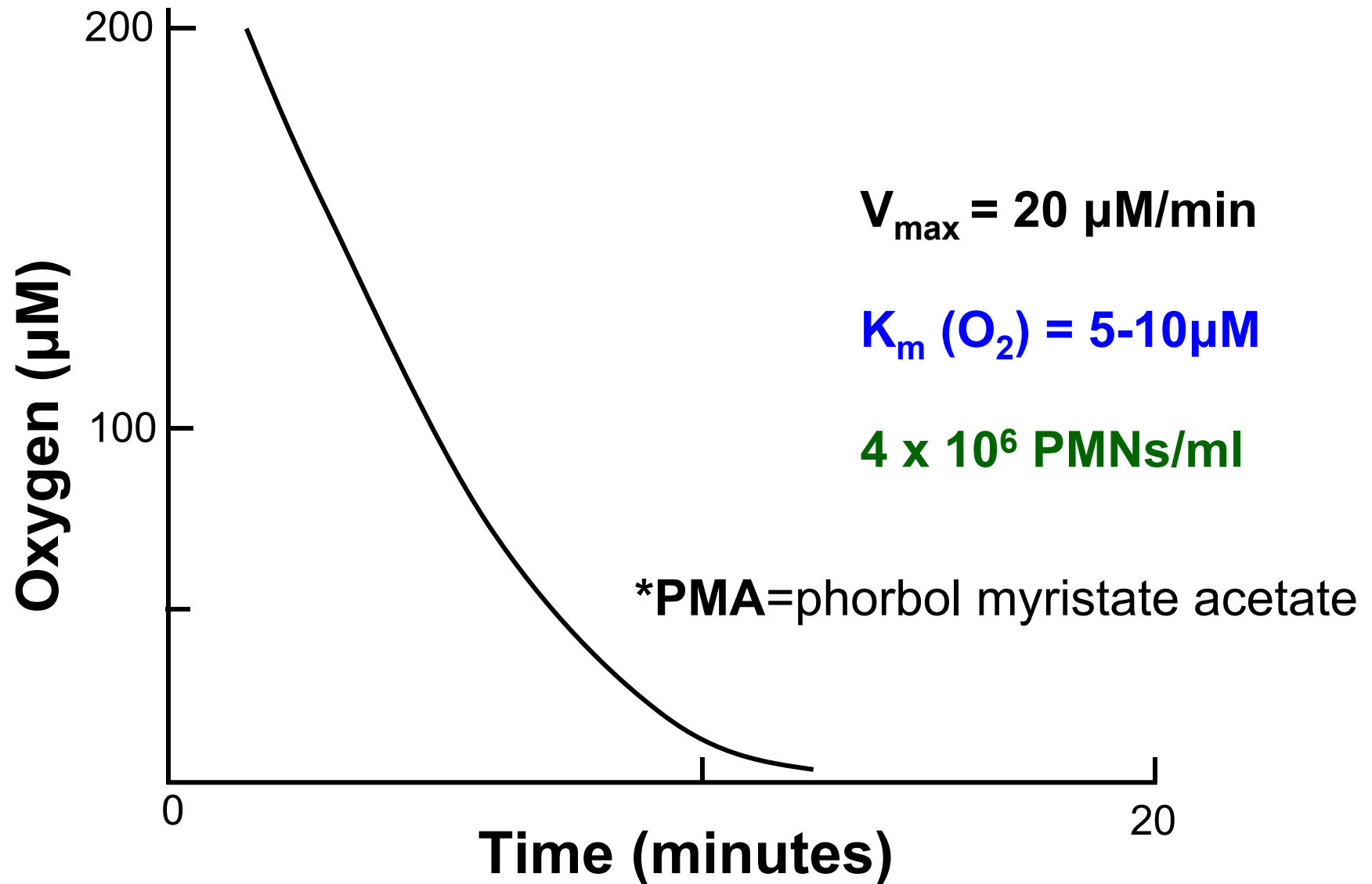
# Chronic Gut Inflammation is Characterized By the Infiltration of Large Numbers of PMNs



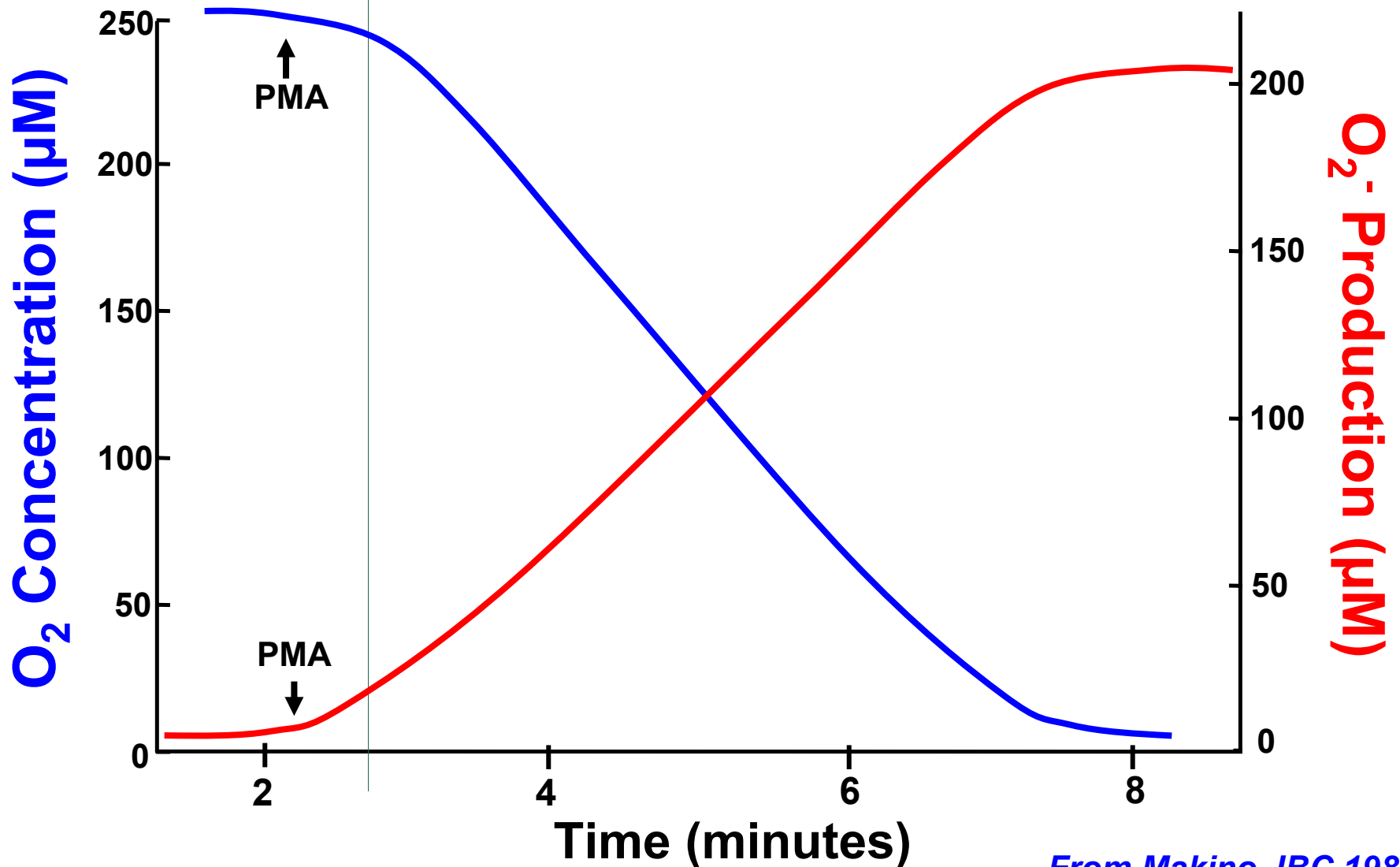
# ROS Production by *Activated* PMNs



# Oxygen Consumption by PMA\*- Activated PMNs in Suspension



# Stoichiometric Conversion of **Oxygen** to **Superoxide** By Activated Neutrophils



*From Makino JBC 1986*

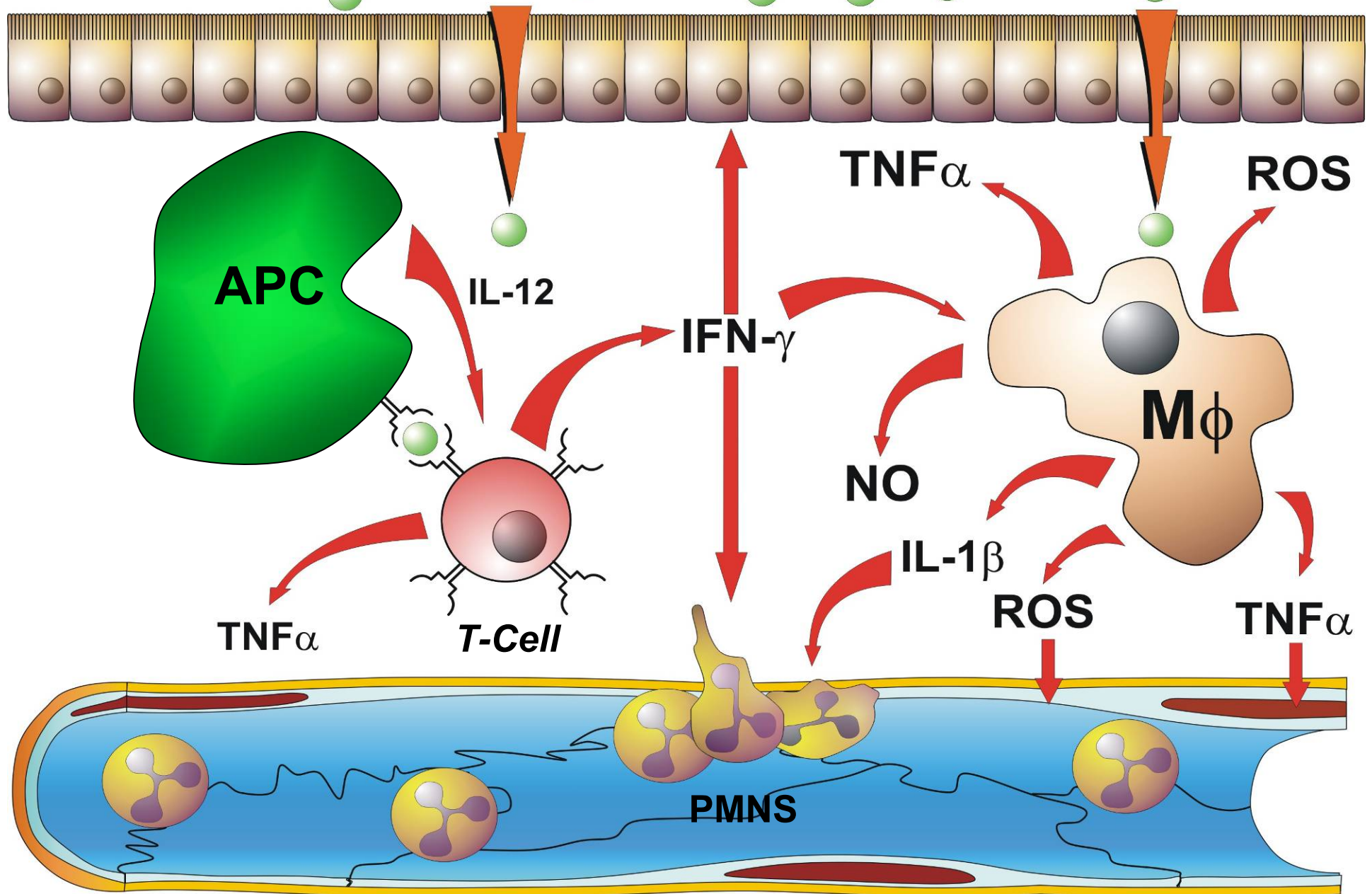
# Reality Check 1

**PMA-Stimulated ROS  
Production by PMNs  
in Suspension:**

***Is this Physiological?***

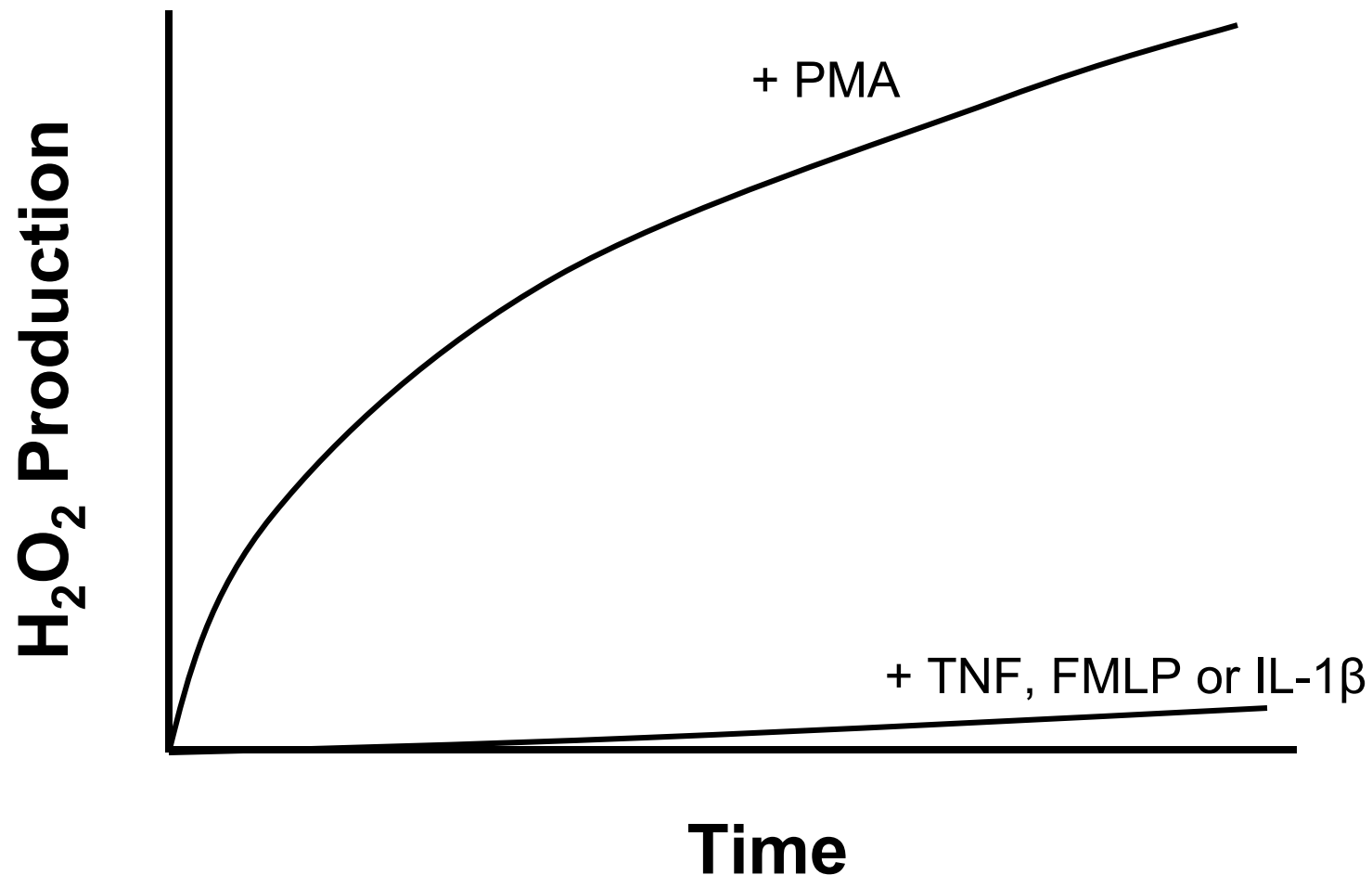


# Inflammatory Mediators (not PMA) Interact with PMNs Associated with Endothelial Cells and the ECM

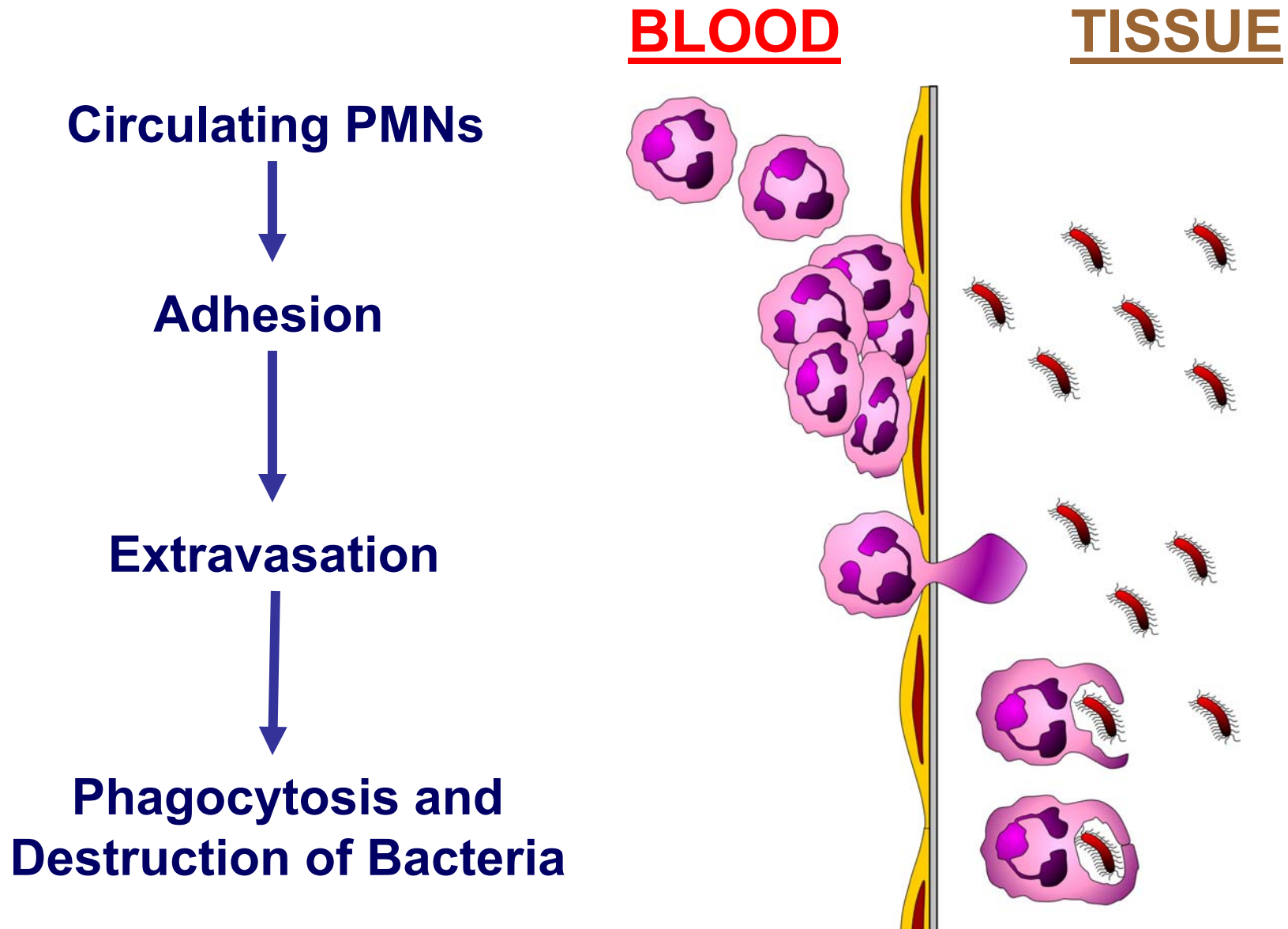




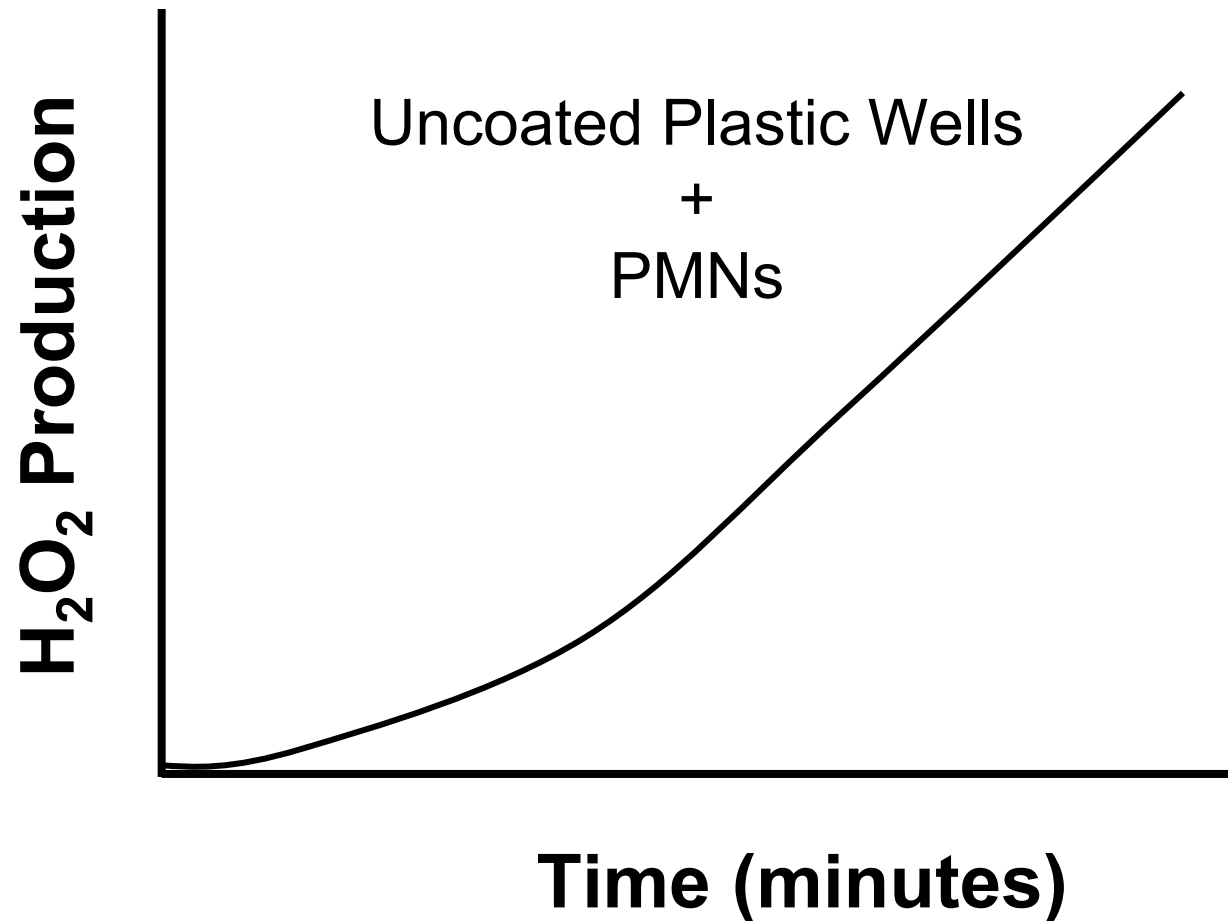
***Biologically-Relevant*** Inflammatory  
Mediators Do **NOT** Activate PMNs *in*  
***Suspension***



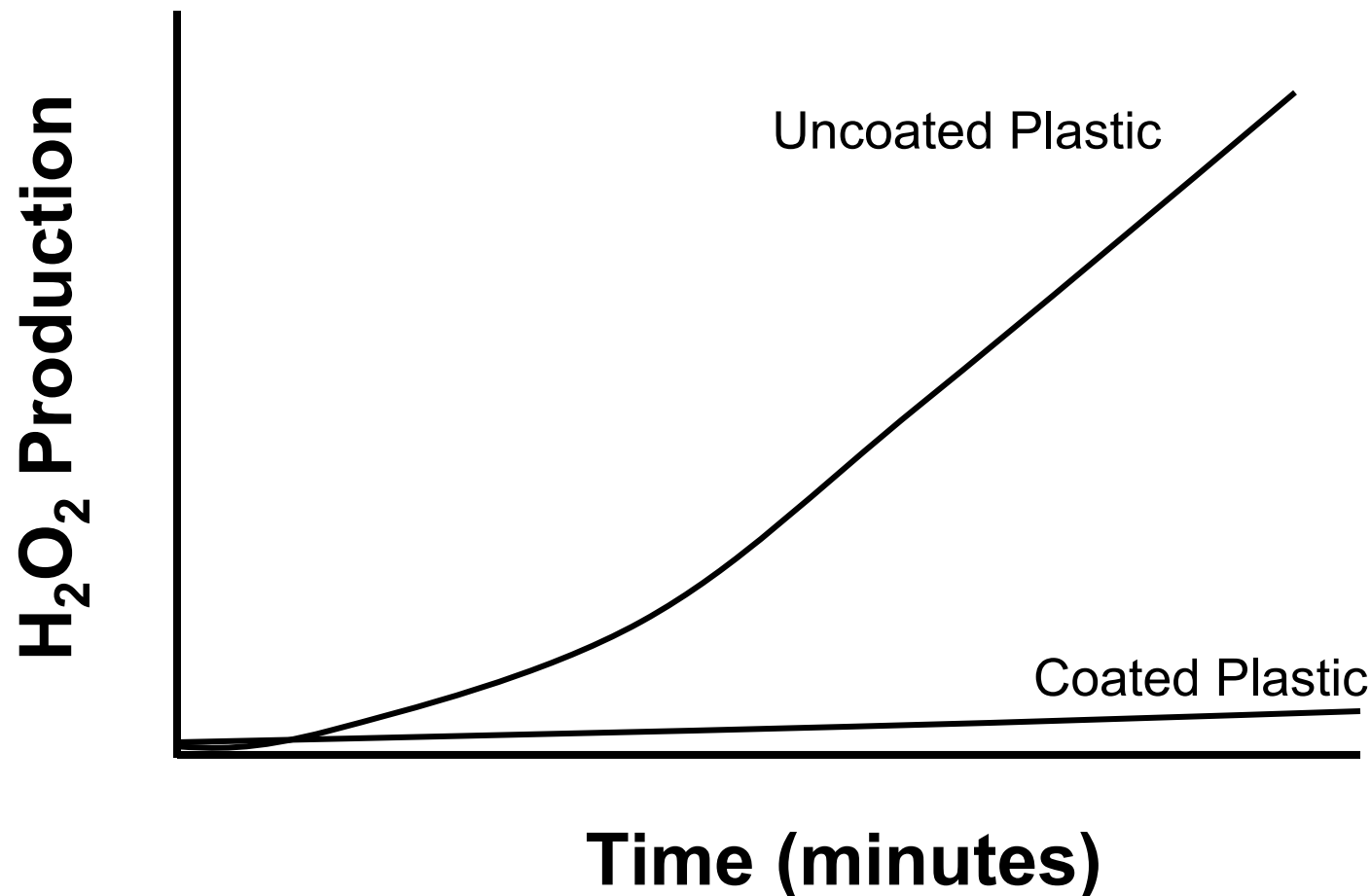
# *PMN-Endothelial Cell* and *PMN-ECM* Interactions In Response to Infection



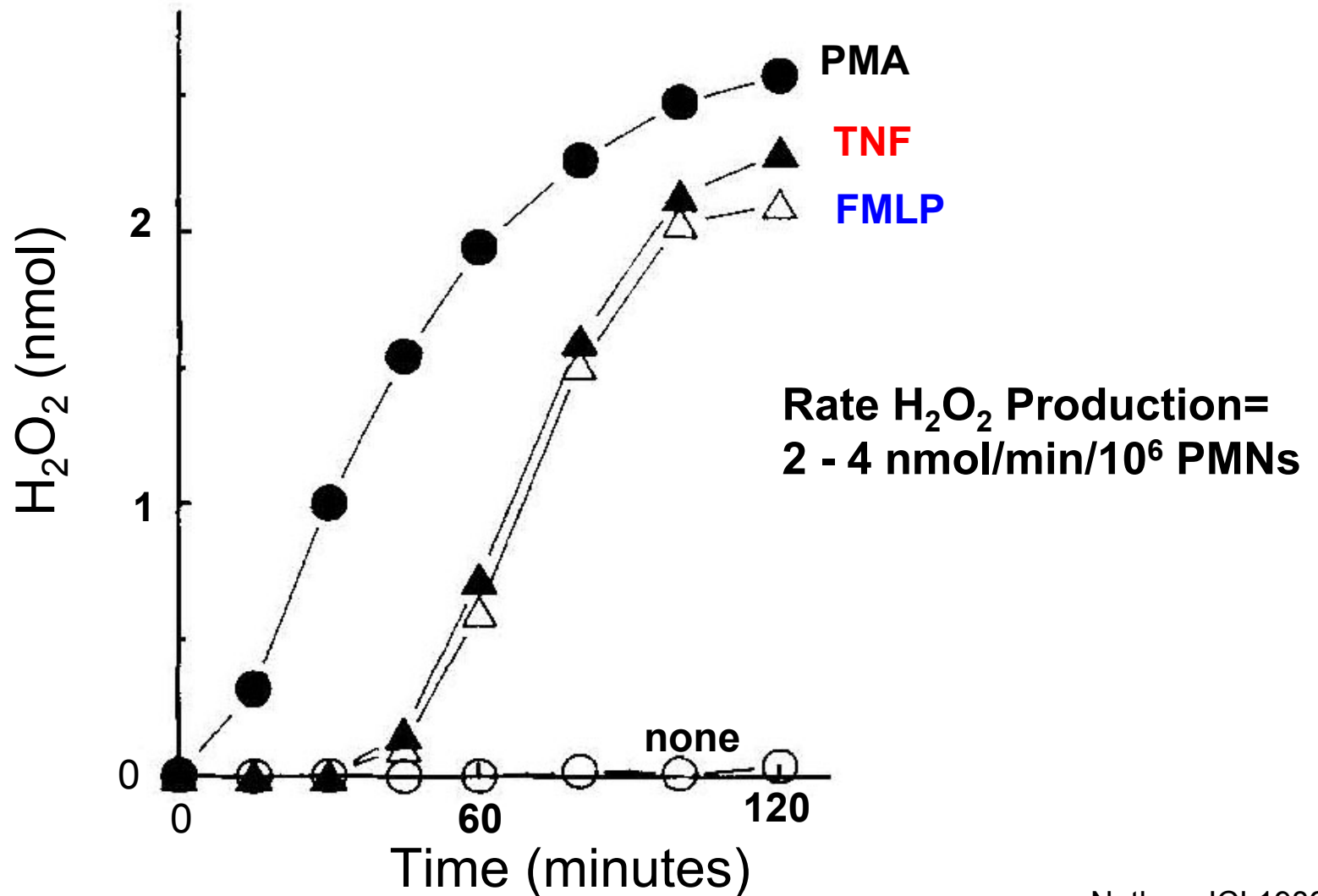
# Plastic Surfaces Activate PMNs



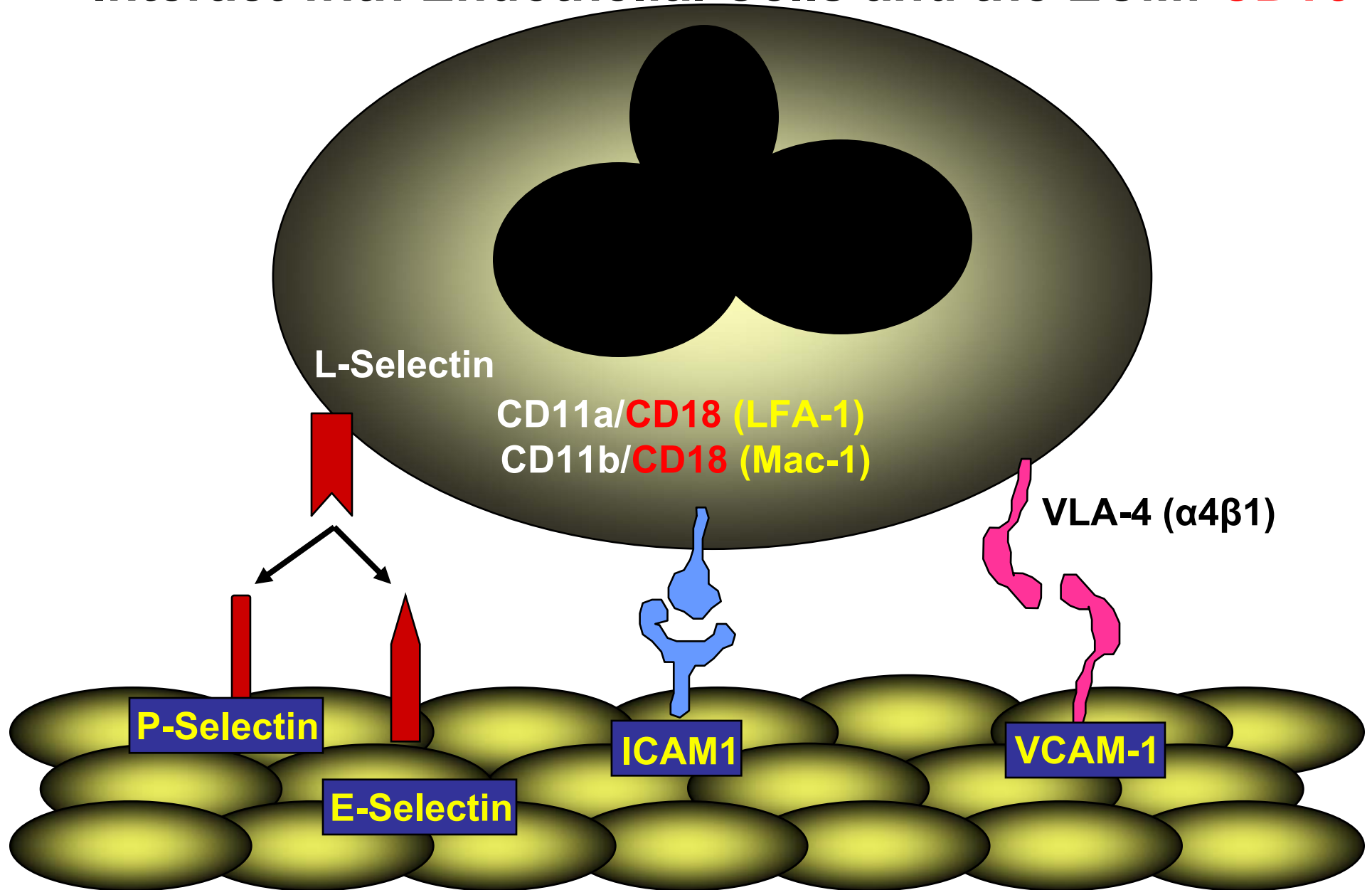
# Coating Plastic Surface with Extracellular Matrix Proteins (Serum, Fibronectin or Laminin) Eliminates Spontaneous Activation of PMNs



# **TNF- $\alpha$** or **FMLP** Induce Large and Prolonged Release of H<sub>2</sub>O<sub>2</sub> When PMNs are Plated on Serum, Fibronectin or Laminin-Coated Plastic Surfaces



# PMNs use a Variety of Adhesion Molecules to Interact with Endothelial Cells and the ECM: **CD18**



# Human and Canine Leukocyte Adhesion Deficiency Disease: *CD18 Deficiency*

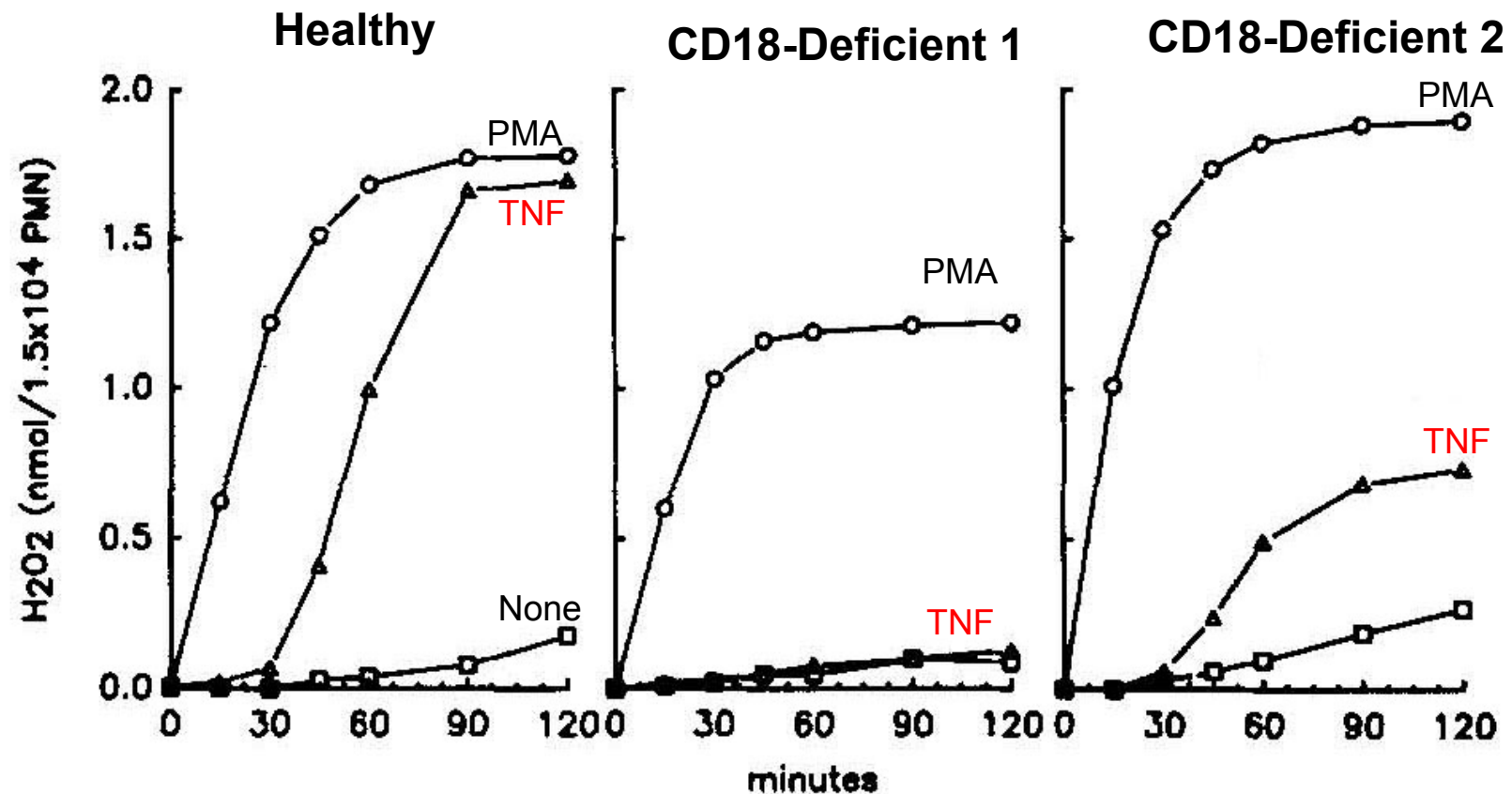


Lymph Node Infection  
(Adenitis): Incision to drain  
abscess took 4 months to  
heal





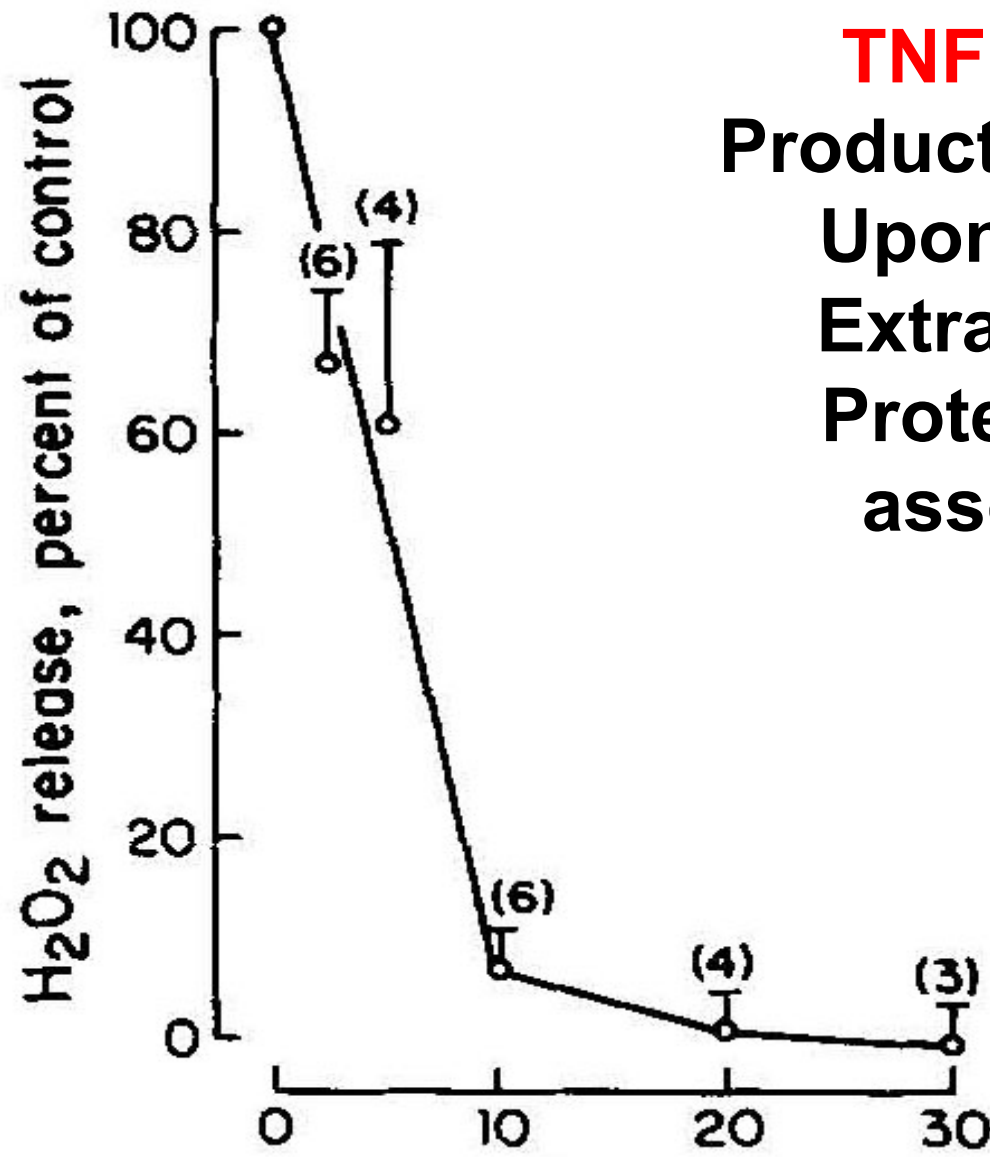
# **TNF-Induced ROS Production is Dependent Upon the Interaction of the $\beta 2$ Integrin (CD18) With ECM Proteins**



Fibronectin-Coated Wells

Nathan JCB 1989

**TNF-Induced ROS  
Production is Dependent  
Upon Interaction of  
Extracellular Matrix  
Proteins with PMN-  
associated CD18**

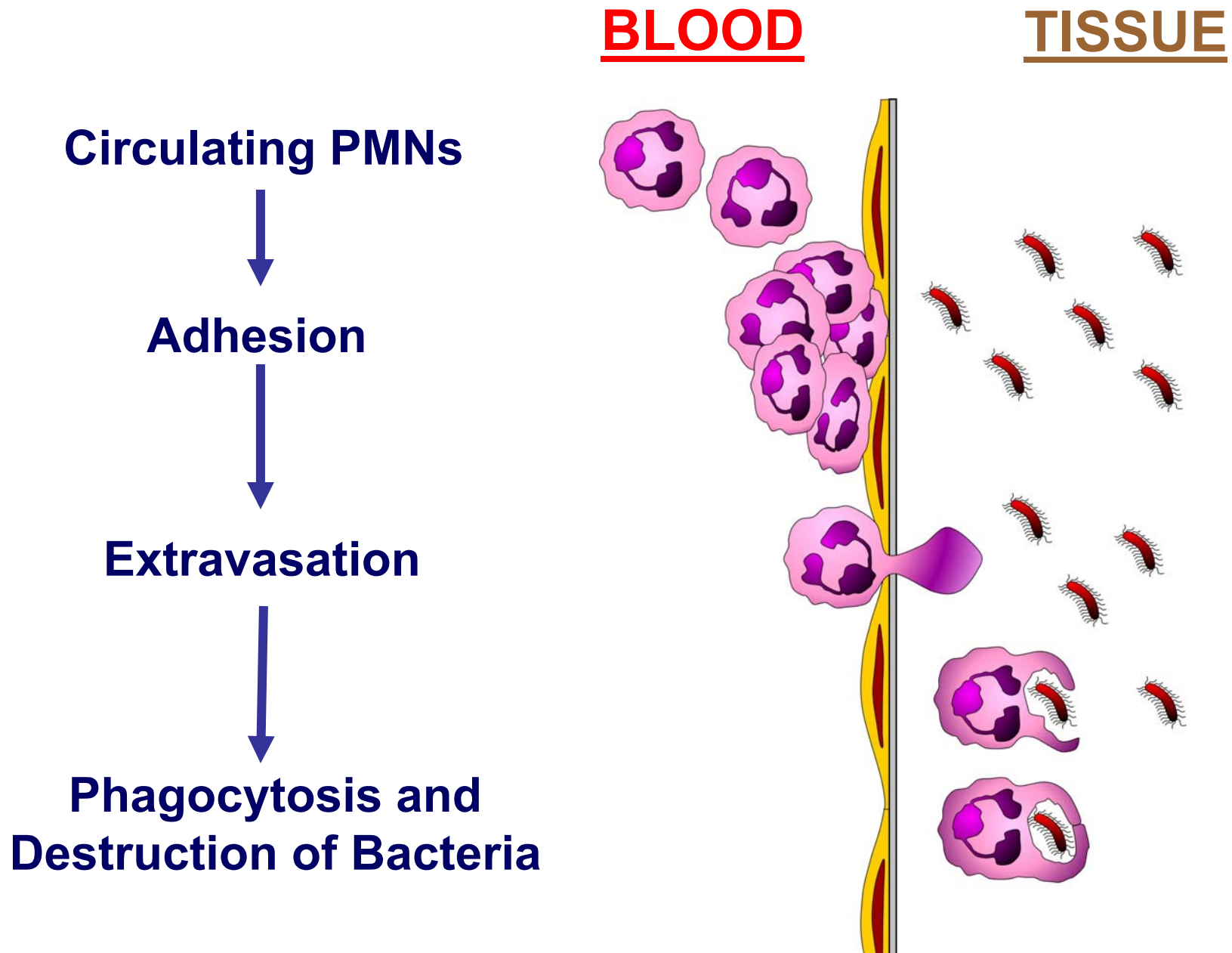


Fibronectin-Coated Wells

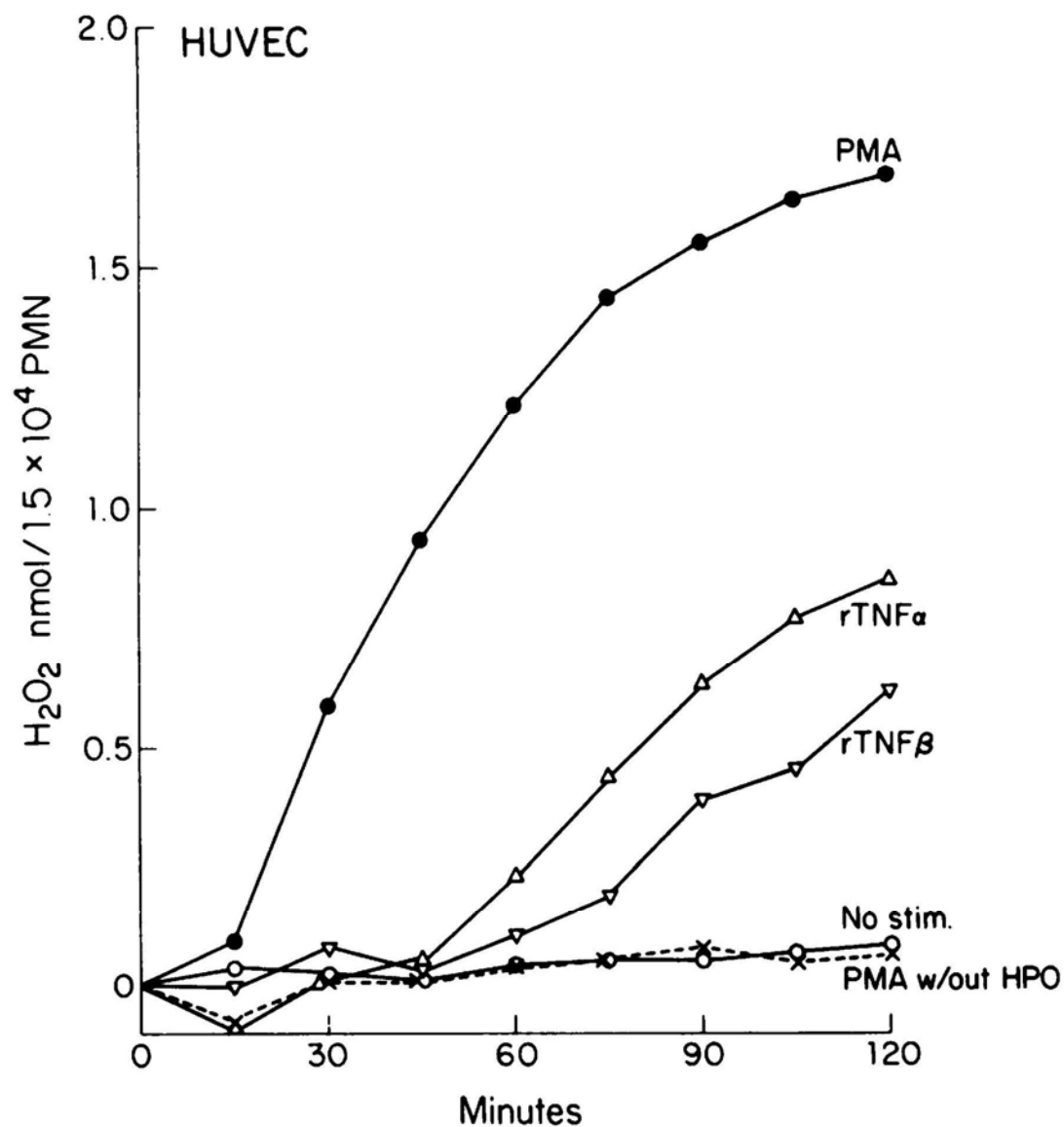
CD18 Ab (ug/ml)

Nathan JCB 1989

# ***Endothelial Cell***- PMN Interactions



# Physiological Levels of TNF Activate PMNs Associated With Endothelial Cell Monolayers



Nathan et. al. 1990

## Reality Check 2

**PMN-Mediated ROS  
Production at Ambient O<sub>2</sub>  
Concentrations**

***Is this Physiological?***

# **Not Really: Consider the Following**

- $pO_2$  is the partial pressure of  $O_2$  in air, solution or in tissue and is expressed in *mm of Hg*. Air contains 21%  $O_2$  which corresponds to a  $pO_2$  of 100 mmHg or 5.2 mM.
- The  $pO_2$  in air-equilibrated buffer (*pH 7.4; 37°C*) is also 100 mmHg (100 Torr) but because of its low solubility in water,  $O_2$  concentration in solution is 200  $\mu\text{M}$ .

# Tissue $pO_2$ is a function of $O_2$ delivery, diffusion distance to nearby capillaries and $O_2$ consumption

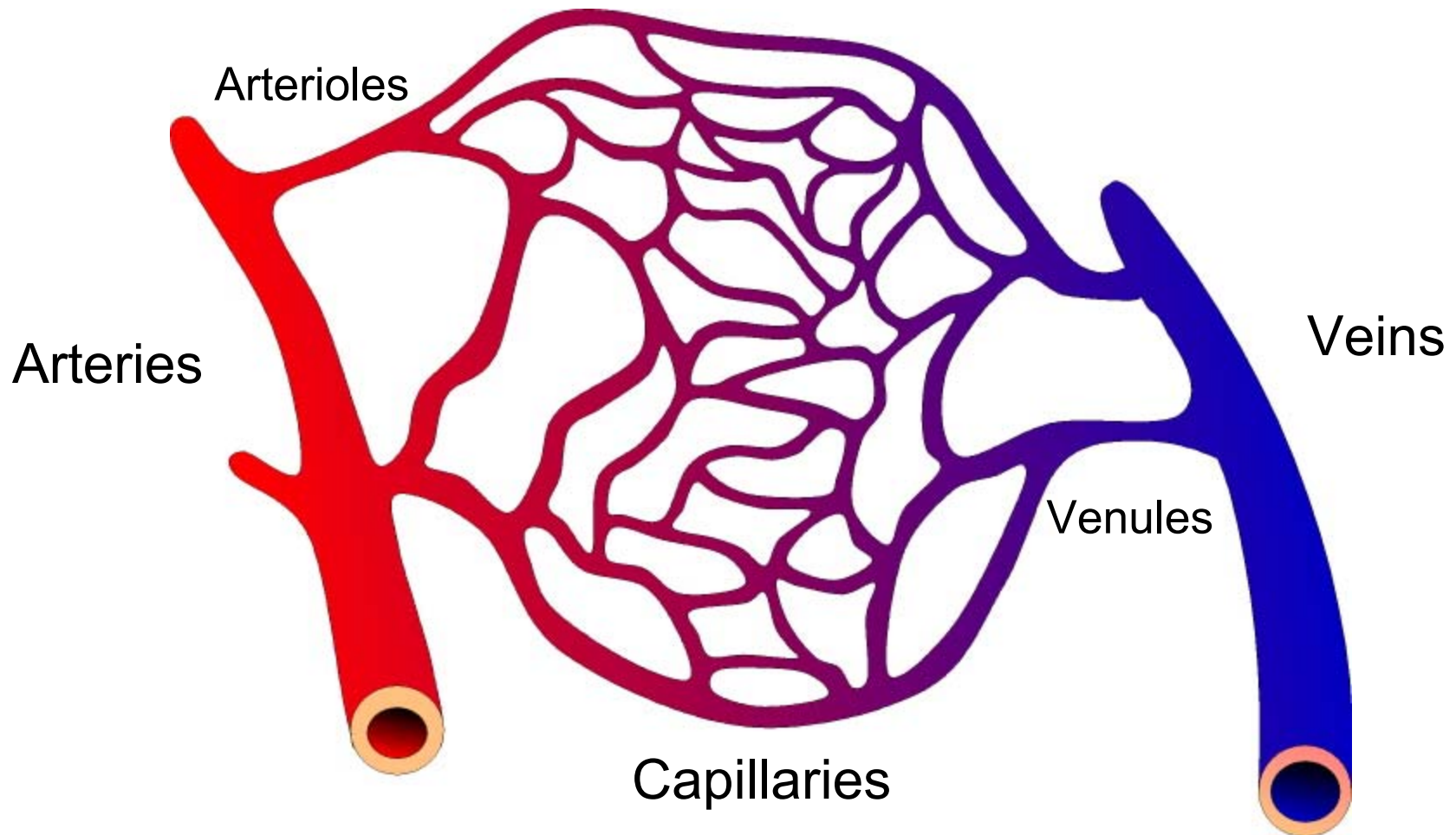
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Except for the *lung, skin epidermis and cornea*, tissue  $pO_2$  ranges from **5-40 mmHg** or from **10-80  $\mu M$**  (assuming an  $[O_2]$  of 100 mmHg or 200  $\mu M$  in solution)

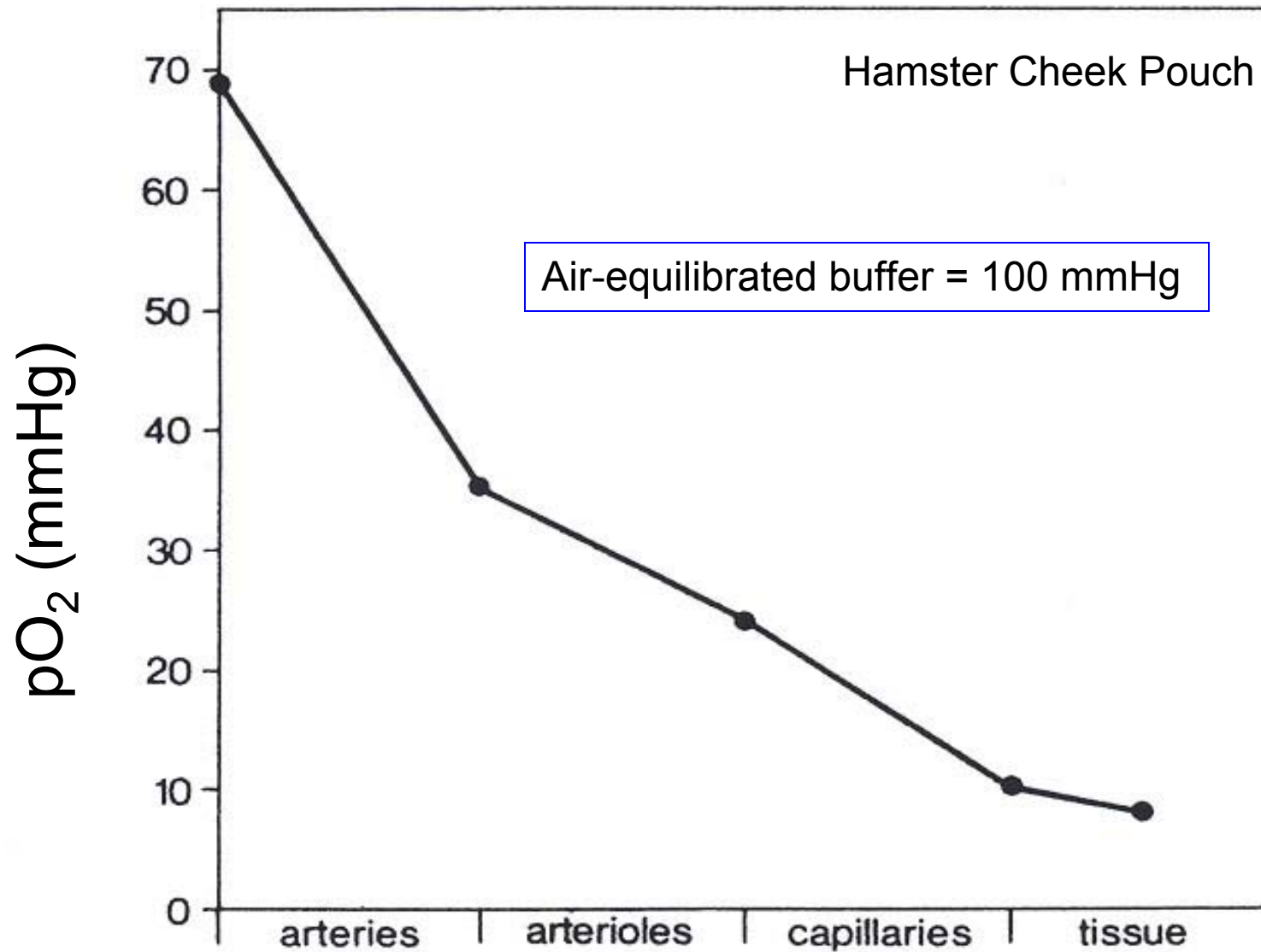
***Tissue  $pO_2$  approximates venous  $pO_2$***



# Inflammation, Tissue Microcirculation and $pO_2$



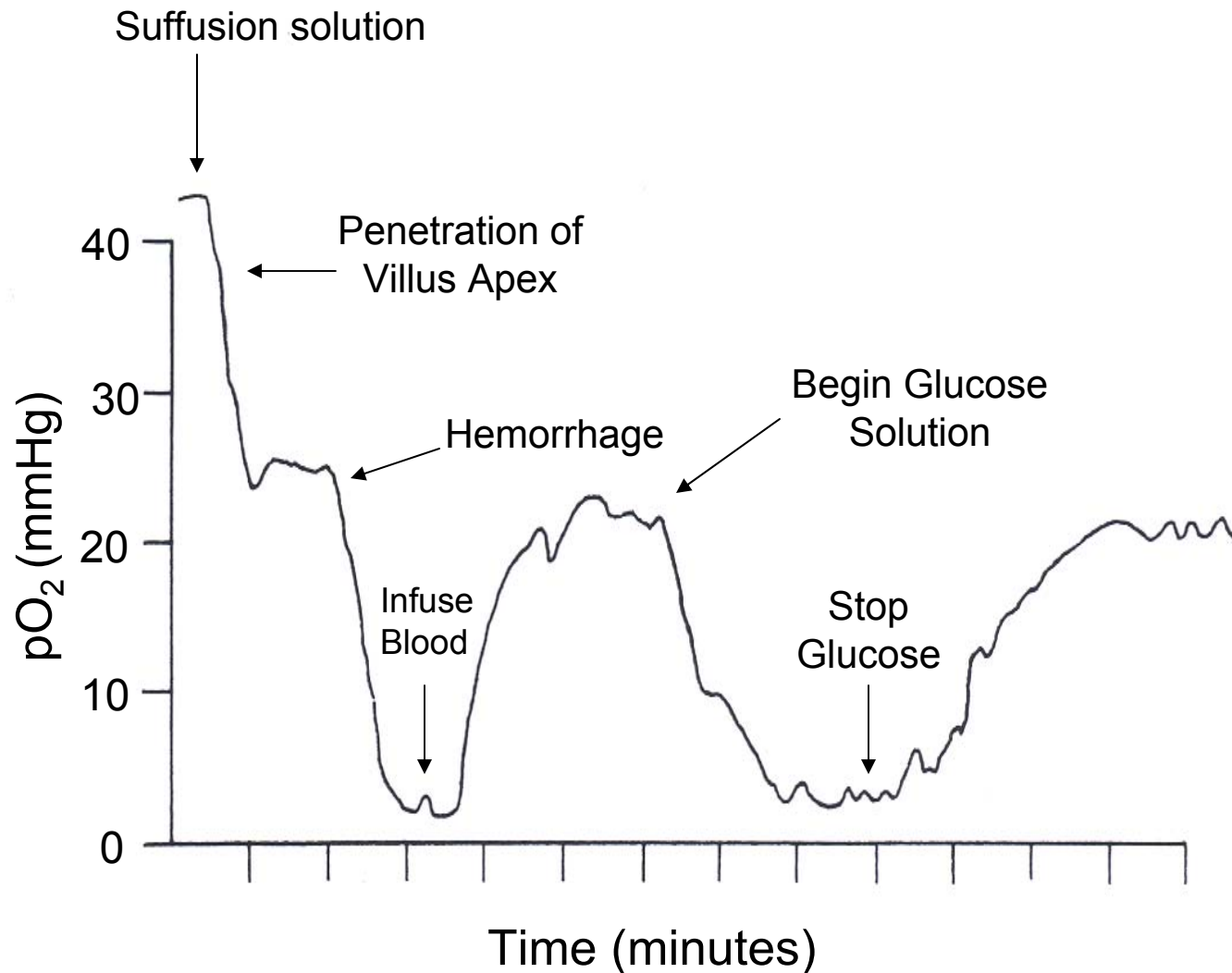
# Perivascular and Tissue pO<sub>2</sub>



# Tissue $pO_2$ Values

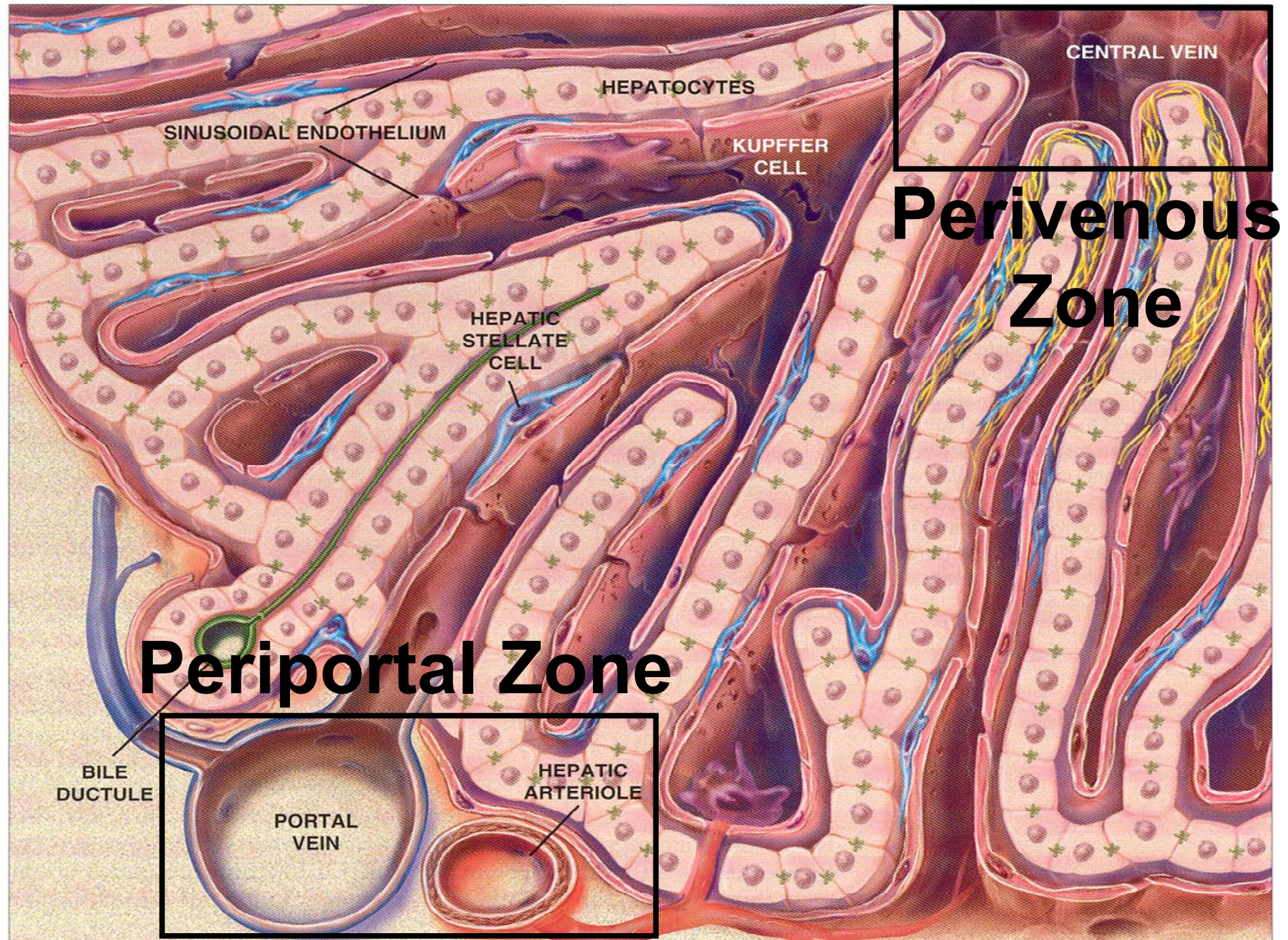
<u>Tissue</u>	<u><math>pO_2</math> (mm Hg)</u>
Kidney (cortex)	45
Kidney (medulla)	23
Skeletal muscle	25
Intestine	25
Brain	15
Liver (Periportal zone)	45
Liver (Perivenous zone)	22

# Physiological $pO_2$ of the Intestine at Rest, Following Hemorrhage and During Nutrient Absorption





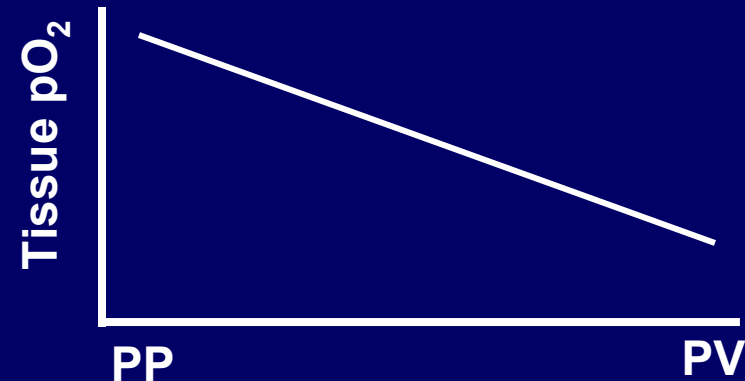
# Oxygen Gradient within the Liver





# Metabolic Heterogeneity of Hepatic Parenchymal Cells along the Sinusoid

*Mitochondrial volume*  
*Oxidative energy metabolism*  
*Oxidative xenobiotic metabolism*  
*Glucose production*



**pO<sub>2</sub> = 45 mmHg (90 μM)**

**Periportal  
Zone 1**

**Perivenous  
Zone 3**

**pO<sub>2</sub> = 22 mmHg (48 μM)**

Glucose uptake  
Glycolysis  
Xenobiotic  
biotransformation

# Why are these considerations important in the context of ROS generation?

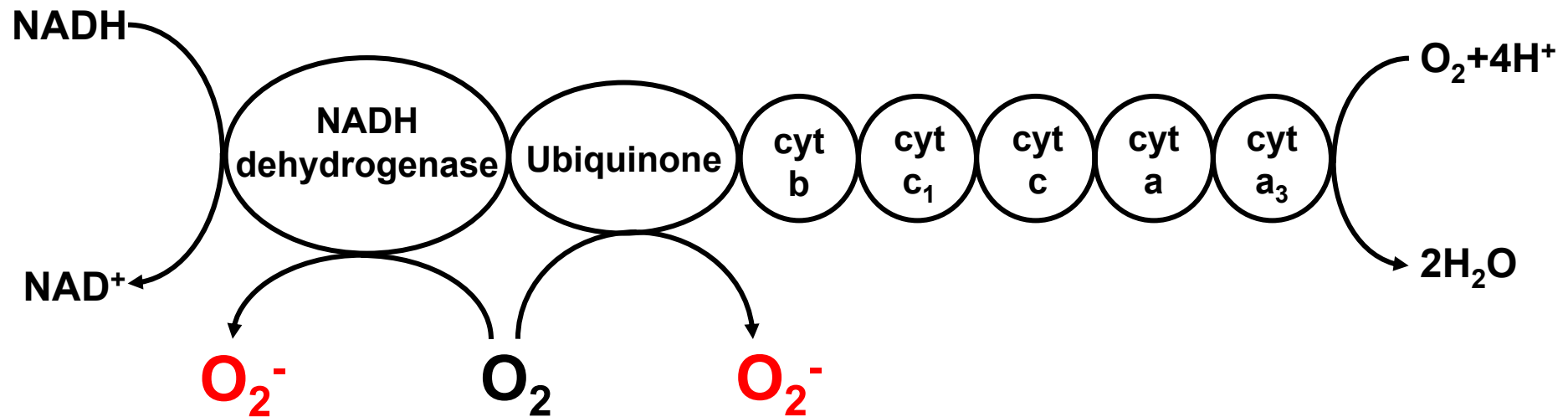
Because most cells and tissue are not exposed to ambient O<sub>2</sub> tension (i.e. 100 mmHg; 200μM). In reality, *physiological pO<sub>2</sub>* for the vast majority of cells and tissue is far below ambient oxygen tension.

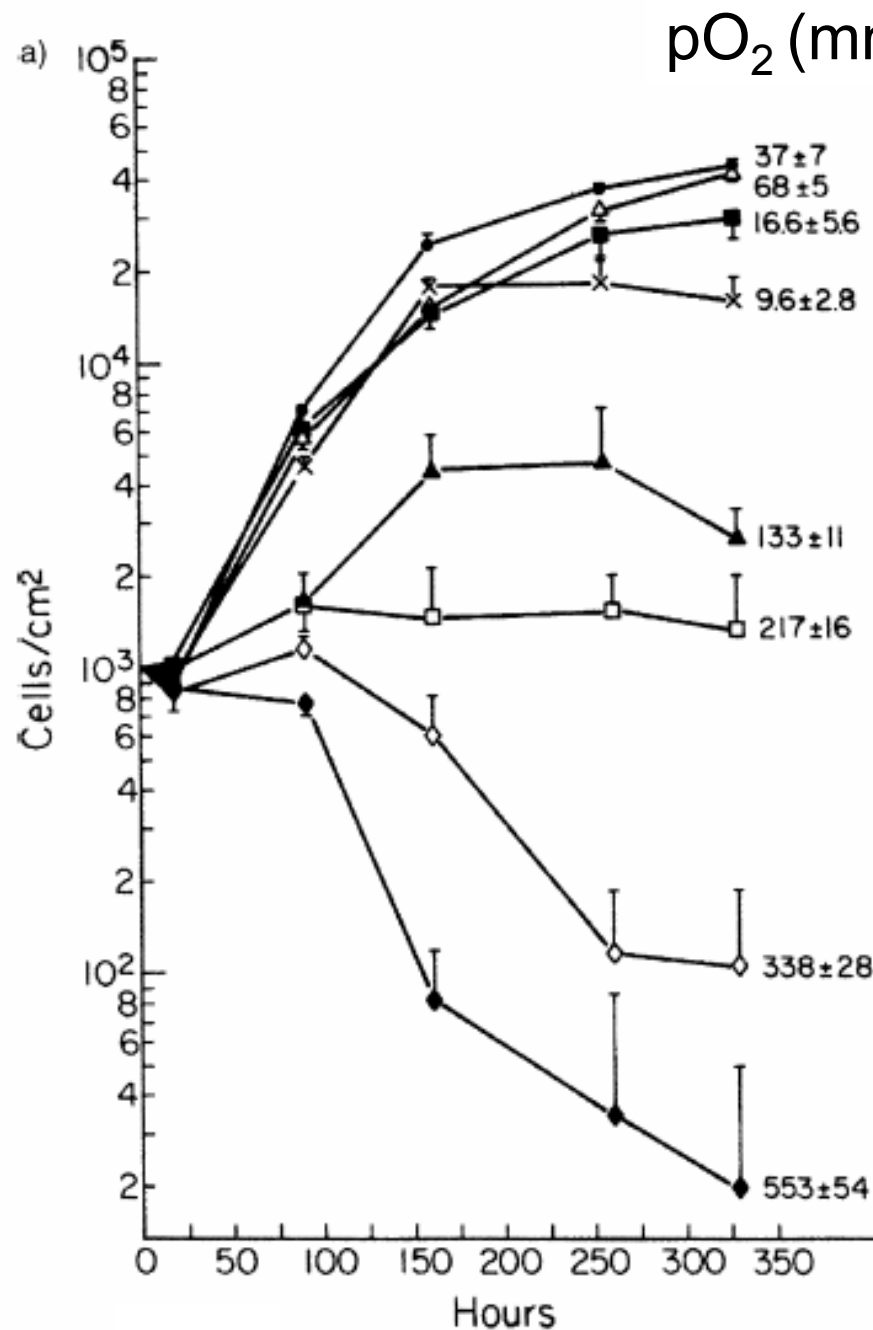


# Cultured Cells Grown at Ambient $pO_2$ are Subjected to *Hyperoxia*

$$dO_2^-/dt = k [O_2] [\text{electron donor}]$$

Doubling of the  $pO_2$  will double  $O_2^-$  production

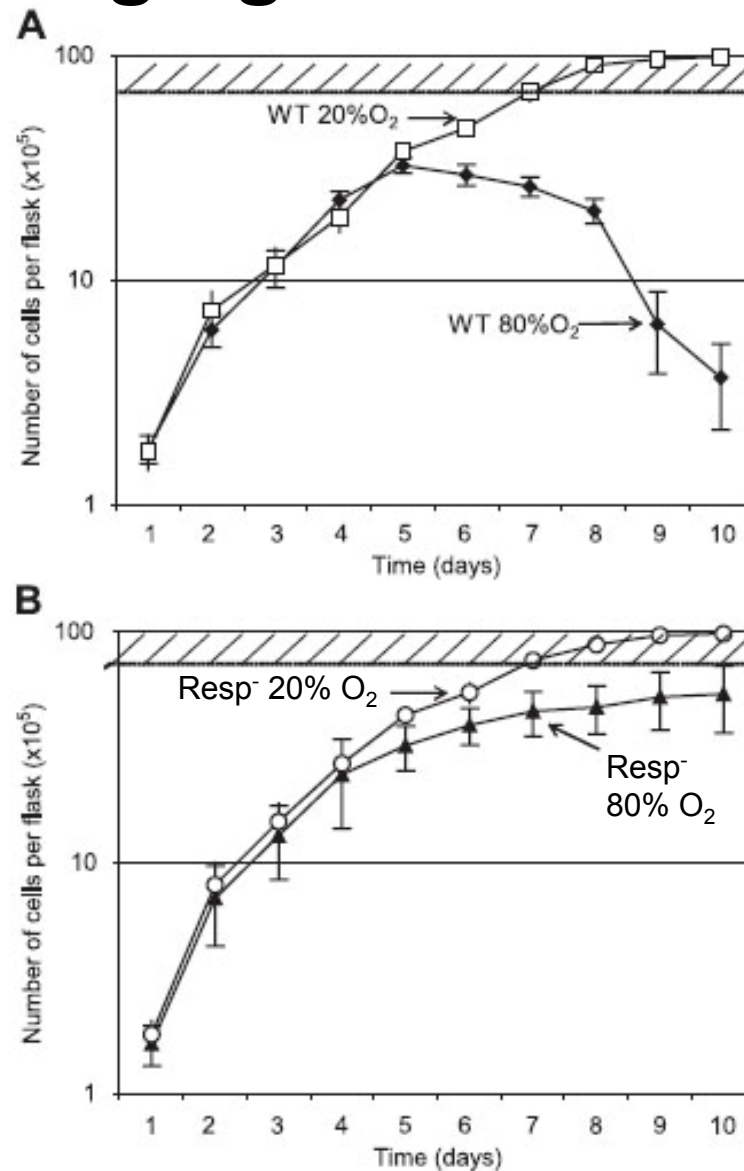




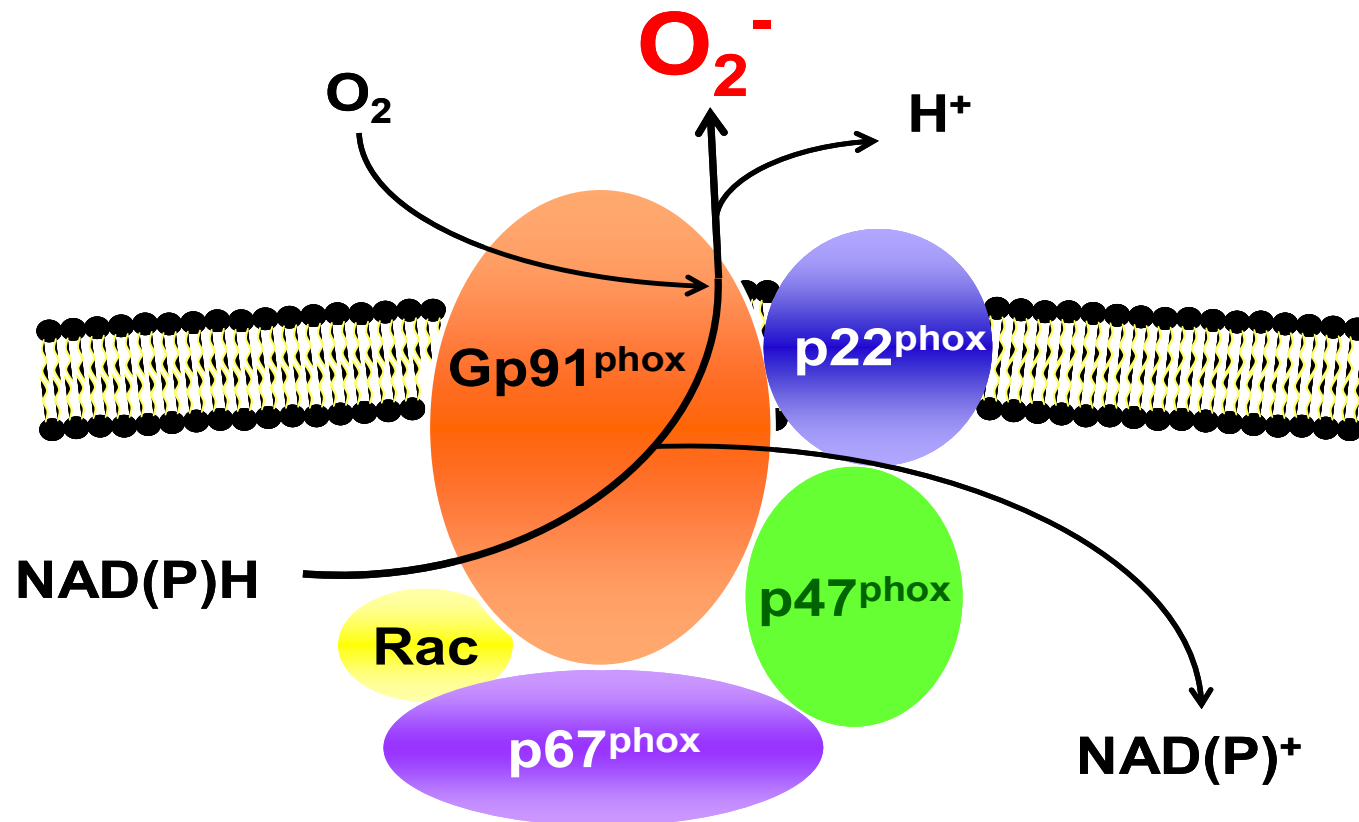
# Oxygen Modulates The Growth of Fibroblasts

Balin and Pratt  
*In Vitro Cell Dev Biol*  
2002

# Respiration-Deficient Cells are Resistant to Damaging Effects of Hyperoxia



# Cultured Cells Grown at Ambient $pO_2$ are Subjected to *Hyperoxia*

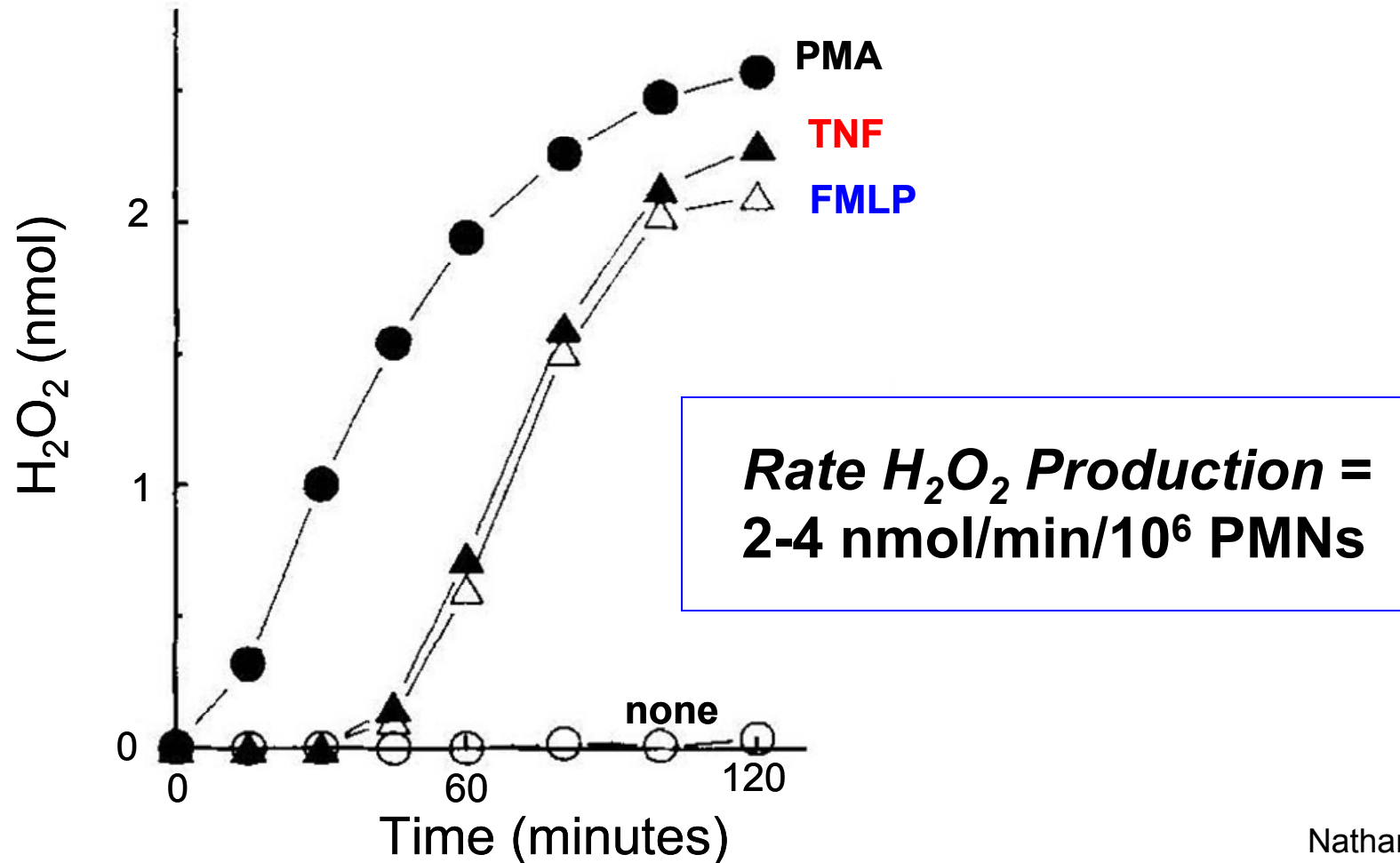


# What are the Rates of PMN-Derived ROS Generation at Physiological $pO_2$ ?

**UNKNOWN.** *However one would predict that  $10^6$  fully-activated PMNs would be functional at  $pO_2$  values as low as 10 mmHg (20  $\mu M$ ) assuming a  $K_m O_2$  of 5-10  $\mu M$  for NADPH oxidase.... However:*

## Reality Check 3

**Rates of ROS Formation by PMNs Would  
Predict That all Tissue  $O_2$  Would be  
Consumed Within A Matter of a Few Minutes**



***Not Necessarily:* Inflammatory Vasodilators (PGE<sub>2</sub>, NO, adenosine) Will Promote the Delivery of O<sub>2</sub> to the Inflammatory Site via Enhancing Tissue Blood Flow** (*O<sub>2</sub> Delivery = Blood Flow x [O<sub>2</sub>]*)

**Poiseuille's Law:** Relates pressure (P), flow (*Q*) and resistance (*R*)

$P = Q \times R$  where  $R = 8L\eta/\pi r^4$       L=length,  $\eta$ =viscosity,  
*r=radius*

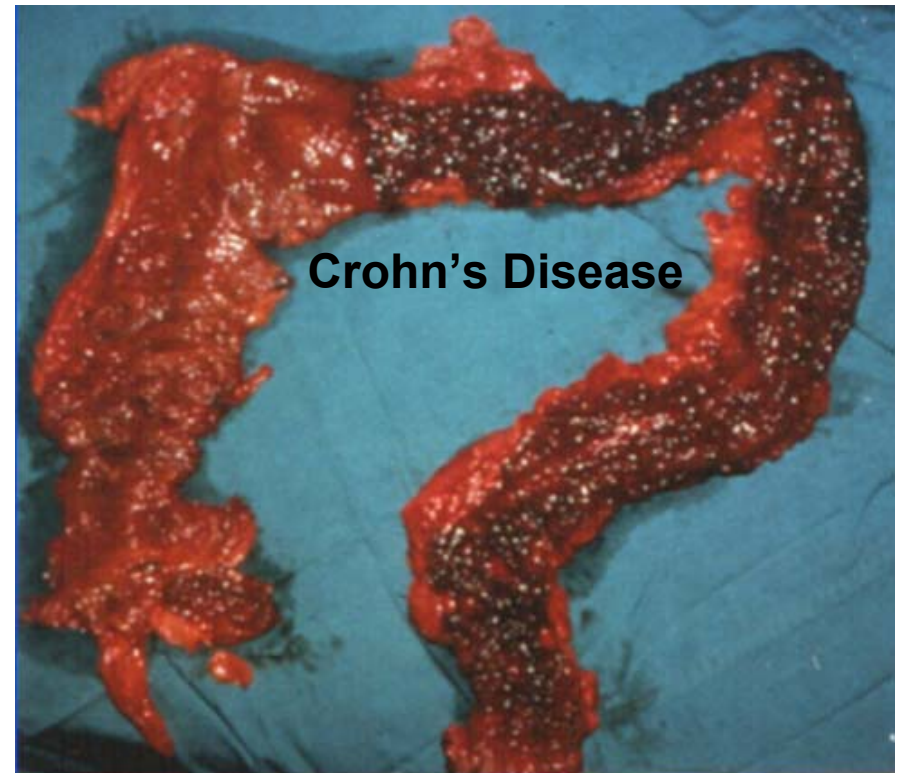
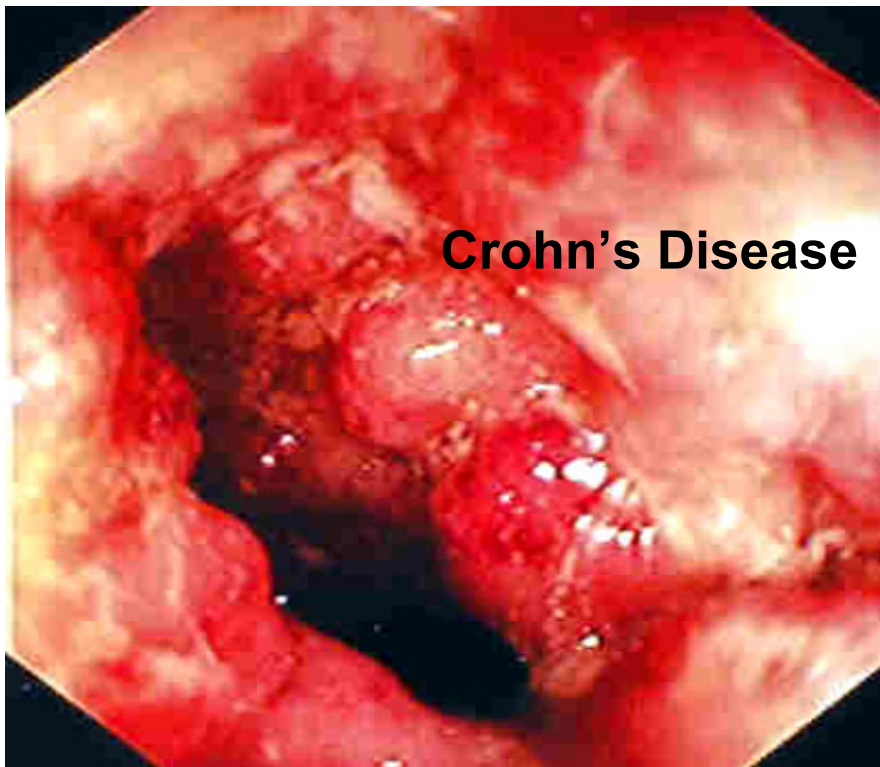
$$Q = \frac{(P)(\pi r^4)}{8L\eta}$$

Note that 2-fold increase in vessel diameter results in a 16-fold increase blood flow

***Inflammation-induced  
hyperemia*** is a good thing  
when responding to a  
bacterial infection or  
tissue injury.



**But what about sustained increases  
in blood flow associated with  
*chronic inflammation?***



**Adding Fuel to the Fire:  
*Potential for Hyperoxic Insult***

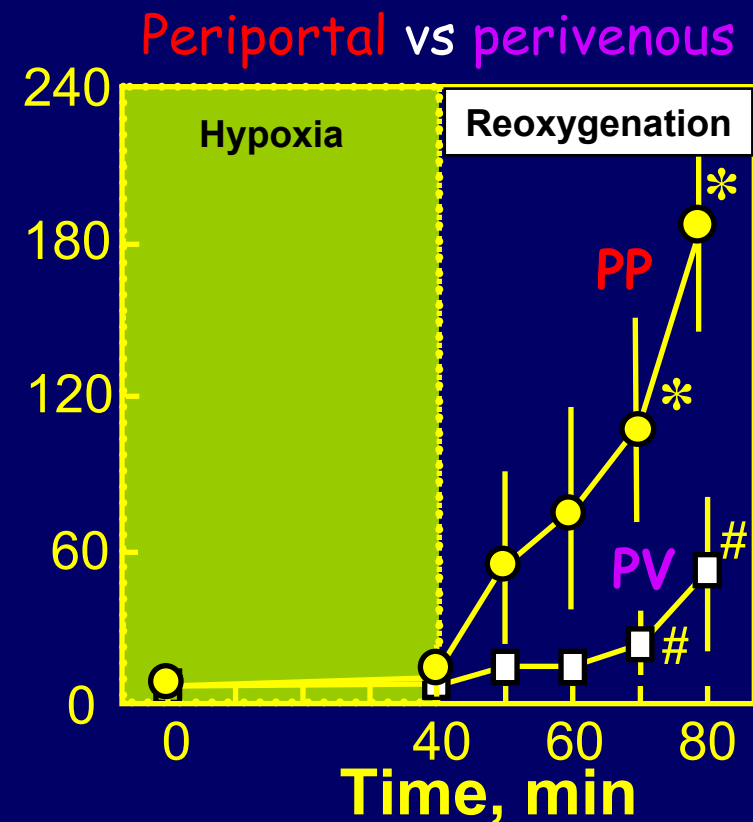
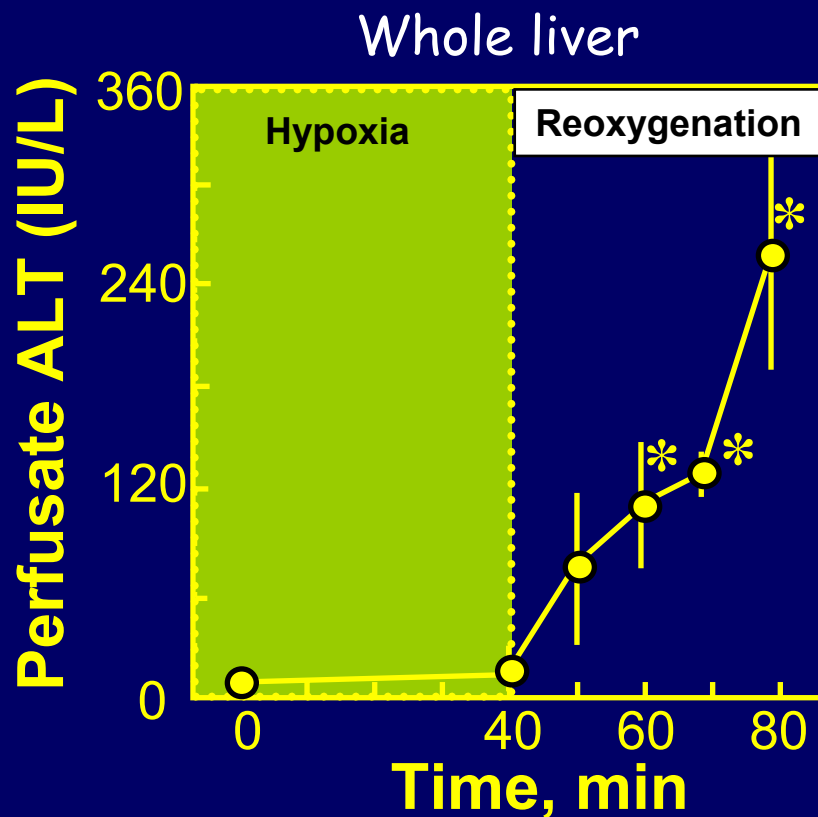
# Summary and Conclusions

- Inflammatory mediator (TNF- $\alpha$ , IL-1 $\beta$ )-induced ROS production by PMNs is dramatically enhanced by the interaction of PMN-associated CD18 with extracellular matrix (ECM) proteins.
- Except for the lung, tissue pO<sub>2</sub> is far below ambient pO<sub>2</sub> and approximates venous oxygen tension ( $\approx$  40 mmHg).
- Blood flow affects tissue pO<sub>2</sub> and thus may regulate the magnitude and duration of PMN-derived ROS generation during times of infection and inflammation.

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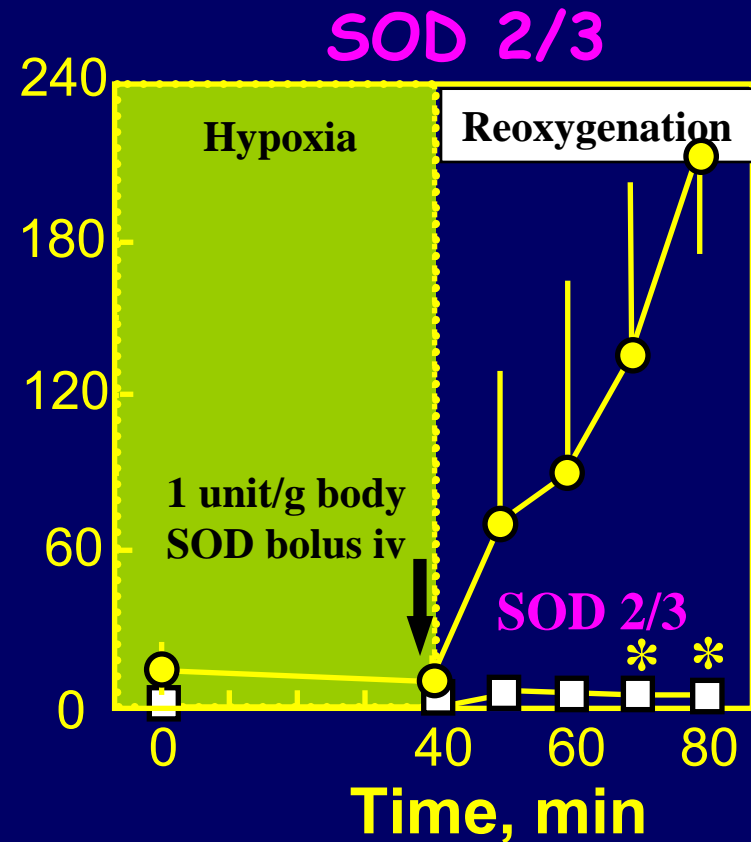
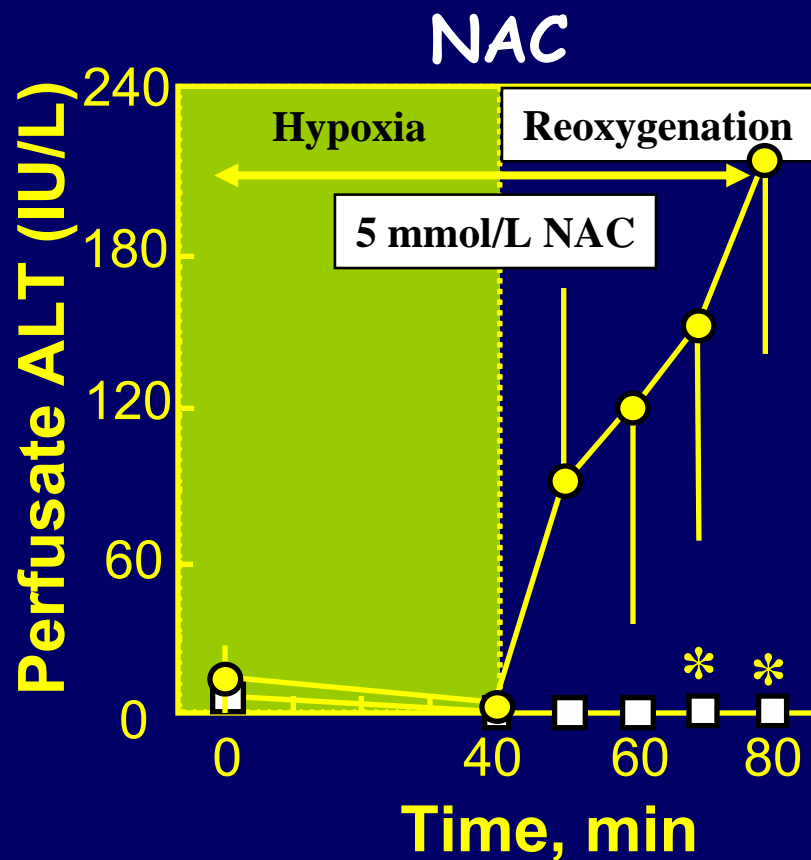
**Physiological fluxes of PMN-derived ROS are dependent upon the physical association of these phagocytes with endothelial cells and ultimately the ECM as well as tissue pO<sub>2</sub>**

# Susceptibility of **Periportal** vs **Perivenous** Zones to Hypoxia/Reoxygenation



AW et. al.

# ROS involvement in HR-induced Periportal injury



Aw et. al.