# Protein oxidation: concepts, mechanisms and new insights

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### **Outline of talk**

- Mechanisms of oxidation of proteins by radical oxidants.
- Mechanisms of oxidation of proteins by non-radical oxidants
- Secondary reactions of reactive protein oxidation intermediates.
- Products of protein oxidation.
- Protein oxidation products as potential markers of disease.

## Reactions of oxidants with biological targets

Oxidants can damage virtually all biological molecules: DNA, RNA, cholesterol, lipids, carbohydrates, proteins and antioxidants.

Extent of damage to particular targets depends on a number of factors including:

concentration of target,
rate constant for reaction of oxidant with target
location of target versus oxidant
occurrence of secondary damaging events
occurrence of transfer reactions
repair and scavenging reactions

Biochem. J., 1997, <u>324</u>, 1-18; Biochim. Biophys. Acta, 2001, <u>1504</u>, 196-219

## Why concentrate on proteins?

### 1) Concentration of target

Proteins are major components of most biological systems:

Organ level (liver, per kg wet weight): 146 g protein, 2.6 g DNA, 49 g total lipid, 3.9 g cholesterol.

Cellular level (per 10<sup>12</sup> leukocytes): 100 g protein, 6.9 g DNA, 8.2 g RNA, 15.6 g total lipid, 2 g cholesterol.

#### Plasma (per dm<sup>3</sup>):

73 g protein, 0.4 g free amino acids, 0.5 g total lipid, 1 g carbohydrates, 1.5 - 2.5 g cholesterol.

Low-density lipoproteins (molecules per particle):

1 protein (4535 amino acids), 1600 cholesterol esters, 700 phospholipids, 600 free cholesterol, 26 free fatty acids, 9 tocopherol.

## Why concentrate on proteins?

### 2) Rate constants for reaction

Rate constants for reaction of HO? with macromolecules:

DNA  $8 \times 10^8 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ 

RNA  $1 \times 10^9 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ 

Hyaluronan  $7 \times 10^8 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ 

Linoleic acid  $9 \times 10^9 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ 

Collagen  $4 \times 10^{11} \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ 

Albumin  $8 \times 10^{10} \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ 

#### **Antioxidants:**

Ascorbate  $1 \times 10^{10} \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ ,

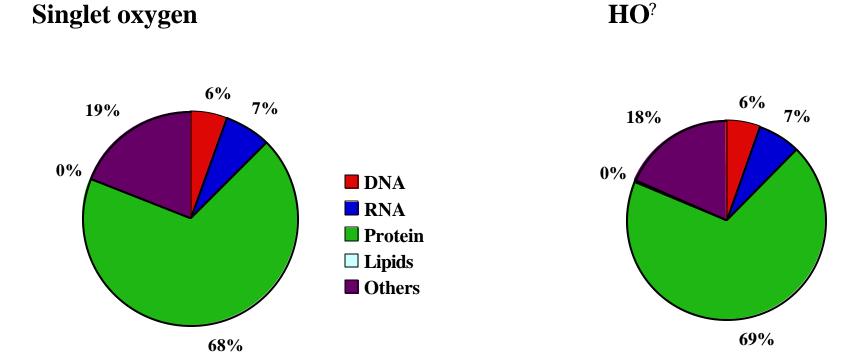
GSH  $1.4 \times 10^{10} \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ ,

Trolox C  $6.9 \times 10^9 \text{ dm}^3 \text{ mol}^{-1} \text{ s}^{-1}$ 

## Why concentrate on proteins?

Kinetic and abundance data can be used to <u>predict</u> sites of damage.

For leukocytes:

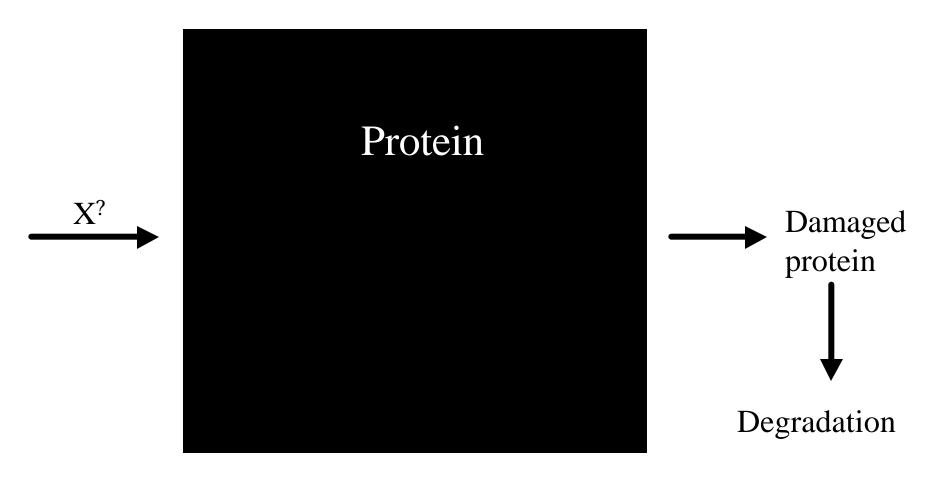


Such data needs to be treated with great caution!

## Protein versus DNA versus lipid oxidation

- Medline / Pubmed searches
  - DNA oxidation 8894
  - Lipid peroxidation 26041
  - Protein oxidation 5463
- Oxidation of proteins studied to a much lesser extent than other targets

## Protein oxidation in the 1960's



One radical = one damaged amino acid residue

## Why is protein oxidation the poor cousin?

Not due to a late start - first reports around 1900

First volume of J. Biol. Chem.

Dakin, H.D. (1906) "The oxidation of amido-acids with the production of substances of biological importance" J. Biol. Chem., 1, 171-176.

Follow-up papers in 1908, occupied approximately half of the total page-count of J. Biol. Chem. for the entire year.

## Why is protein oxidation the poor cousin?

- Complexity of targets ?
- Complexity of products?
- Complexity of mechanisms?
- Lack of sensitive, stable, readily detectable markers of damage?

## Sites of oxidant damage on proteins

• Backbone - primarily hydrogen atom abstraction at alpha carbon

$$\begin{array}{c|cccc}
O & R & H & O & R & H \\
\hline
N & H & O & R & H & O \\
R & H & O & R & H & O
\end{array}$$

- can result in backbone fragmentation
- Side-chains 20 different types (excluding unusual amino acids and any post-translational modifications).
  - hydrogen abstraction primarily with aliphatic
  - addition primarily with aromatic
    - usually results in the formation of altered side-chains

Chem Rev, 1987, <u>87</u>, 381-398; Free Rad Biol Med, 1990, <u>9</u>, 315-325; J Biol Chem, 1987, <u>262</u>, 9895-9920

## Selectivity of damage by different oxidants

• The most reactive radicals tend to be the least selective

e.g HO? - difference in rate constants is relatively small

Most reactive: Trp, Tyr, His, Met, Cys, Phe, Arg: k ca. 10<sup>10</sup> dm<sup>3</sup> mol<sup>-1</sup> s<sup>-1</sup>

Least reactive: Ala, Asp, Asn: k ca.  $10^7 - 10^8 \,\mathrm{dm}^3 \,\mathrm{mol}^{-1} \,\mathrm{s}^{-1}$ 

Result - most side-chains are oxidised

Reaction with backbone sites k ca.  $10^9$  dm<sup>3</sup> mol<sup>-1</sup> s<sup>-1</sup>

Significant backbone fragmentation as well as side-chain oxidation

- Less reactive radicals tend to be more selective
  e.g. CCl<sub>3</sub>OO? difference in value of rate constants between most reactive side-chain (Trp k ca. 9 x 10<sup>7</sup> dm³ mol<sup>-1</sup> s<sup>-1</sup>) and least (aliphatic side-chains no measurable reaction) very large.
- Many radicals are electron-deficient (electrophilic) and hence react most rapidly with electron-rich side-chains (Trp, Tyr, His, Met, Cys, Phe). Few nucleophilic oxidants (e<sup>-</sup>, Ph<sup>?</sup>, CO<sub>2</sub><sup>?-</sup>).

### Sources of kinetic and associated data

Compilations of kinetic data:

Website: NDRL/NIST Solution Kinetics Database - 14,000 rate constants

http://www.rcdc.nd.edu/RCDC/RadChemHomePage.html

 Reduction potentials for one-electron reactions involving radicals
 J. Phys. Chem. Ref. Data, 1989, <u>18</u>, 1637-1755.

http://www.rcdc.nd.edu/RCDC/RadChemHomePage.html

# Selectivity of damage between different sites by a single oxidant

- Kinetic data does <u>not</u> usually yield information on selectivity of damage at different sites, unless specific absorptions are monitored - usually only possible for aromatic and sulfurcontaining residues.
- Number of factors influence which sites are most favored
  - Stability of incipient radical (tertiary > secondary > primary; delocalisation on to other atoms)
  - Statistics number of available C-H bonds / sites of addition
  - Accessibility (buried versus exposed; steric and charge interactions)

## Backbone fragmentation induced by radicals

#### Common intermediate implicated in majority of mechanisms

- Formation of?-carbon radical via direct, or indirect reaction detection by EPR spin trapping (e.g. Chem. Res. Toxicol., 2000, <u>13</u>, 1087-1095).
- Subsequent formation of peroxyl radical in presence of  $O_2$ .
- Little backbone fragmentation in absence of  $O_2$ .

# Mechanisms of side-chain oxidation by radicals: aliphatic residues

Hydrogen atom abstraction gives common intermediates but

ratio of intermediates formed at different C-H positions varies with attacking radical.

centered radical

Direct, rapid-flow, EPR spectroscopy studies can give information on selectivity of initial radical attack for amino acids and peptides.

centered radical

J. Chem. Soc., Perkin Trans. 2, 1998, 2617-2622; Biochim. Biophys. Acta, 2001, 1504, 196-219

# Mechanisms of side-chain oxidation by radicals: aliphatic residues

Initial carbon-centred radicals undergo rapid reaction with  $O_2$  to give peroxyl radicals. Dimers formed in absence of  $O_2$ .

$$R^? + O_2 \longrightarrow ROO^? \text{ or } R^? + R^? \longrightarrow R-R$$

Fate of peroxyl radicals

- Reaction with another peroxyl radical to give ROO-OOX (X = R, H).
- Hydrogen atom abstraction gives hydroperoxide:
   ROO? + XH → X? + ROOH → ROH + carbonyls

Elimination reactions - special case for Ser, Thr and few other residues

$$-C-OO^?$$
  $\longrightarrow$   $-C=O$  +  $HOO^?$ 

Major products from aliphatic side-chains: peroxides, alcohols and carbonyls.

Biochem J, 1995, <u>305</u>, 643-649; Arch Biochem Biophys, 1996, <u>336</u>, 163-172; Biochem. J. 1997, <u>324</u>, 1-18

## Specific aliphatic side-chain oxidation products

Glutamic acid hydroperoxides Lysine hydroperoxides

alcohols

alcohols

Leucine hydroperoxides carbonyl compounds

alcohols

? -ketoisocaproic acid Proline hydroperoxides

isovaleric acid

isovaleraldehyde 5-hydroxy-2-aminovaleric acid

isovaleraldehyde oxime carbonyl compounds

carbonyl compounds

**Arginine** hydroperoxides

Glycine Aminomalonic acid 5-hydroxy-2-aminovaleric acid

Valine hydroperoxides Isoleucine hydroperoxides,

alcohols

carbonyl compounds carbonyl compounds

**Methionine Methionine sulphoxide** 

Cysteine Cystine,

Oxy acids

alcohols,

Free Radic. Biol. Med., 1999, <u>27</u>, 1151-1163

# Peroxides are major initial products of HO? attack on amino acids, peptides and proteins in the presence of $O_2$

Substrate	% yield of hydroperoxide groups formed from initial HO .
N-Ac-Lys-NH <sub>2</sub>	26
Gly-Lys-Gly	34
Poly-lysine	64
Melittin	16
Protamine	33
Insulin	12
RNase A	53
BSA	36

## Formation of peroxides on proteins

#### Does it matter what the attacking radical is? - NO

High energy radiation (?- or X-rays, UV, visible light with sensitizer)

Metal ions / ascorbate

Metal ions / peroxide systems (HO?, RO?)

Thermo-labile azo compounds  $+ O_2 (ROO^?)$ 

Peroxynitrite

Activated white cells

Hemoprotein / peroxide systems

#### Does it matter what the amino acid / peptide / protein is? - NO

Formed on most amino acids, and <u>all</u> peptides and proteins tested Detected on both isolated proteins and proteins in cells

Yield and exact structure of peroxides formed is dependent on target, the attacking radical,  $O_2$  concentration, presence of reductants / antioxidants / metal ions (etc...)

Redox Rep, 1997, 3, 99-110; Biochem J, 1997, 324, 1-18; Biochim Biophys Acta, 2001, 1504, 196-219

## Fate of protein peroxides

Protein-OOH + 2e<sup>-</sup>

**Protein-OH** 

Protein-OOH + M<sup>n+</sup>

? **Protein-O**? +  $M^{(n+1)+}$  +  $HO^{-}$ 

Protein-OOH + UV / heat ? Protein-O $^{\circ}$  + HO $^{\circ}$ 

**Protein-O?** 

**Protein-C**?

(1,2 hydrogen atom shifts)

**Protein-O?** 

Protein-carbonyl +  $\mathbb{R}^{?}$  (?-scission reactions)

**Protein-C**? + released carbonyl or

Protein- $\mathbb{C}^? + \mathbb{O}_2$ 

**Protein-OO?** 

Protein- $OO^? + RH$ 

? Protein-OOH + R?

Biochem. J., 1995, 305, 643-649; Arch. Biochem. Biophys., 1996, 336, 163-172; Chem. Res. Toxicol, 2000, 13, 1087-1095; Free Radic. Biol. Med., 2002, 32, 1171-1184.

## Transfer of damage within proteins

- Evidence for radical transfer from side-chains sites to backbone via mediation of alkoxyl radicals
  - Favorable process due
- to stability of ? -carbon radical
- relief of steric crowding
- stability of carbonyl product

Detected by Quantified EPR spin by HPLC trapping

- Results in loss of side-chain group as reactive aldehyde / ketone and formation of backbone radical -> backbone cleavage
- Similar reactions with thiyl radical from cysteine?

Chem Res Toxicol, 2000, <u>13</u>, 1087; Free Radic Biol Med, 2002, <u>32</u>, 1171, J Am Chem Soc, 2003, <u>125</u>, 2042

### Occurrence of chain reactions

Number of reactions of protein radicals that give rise to other radicals:

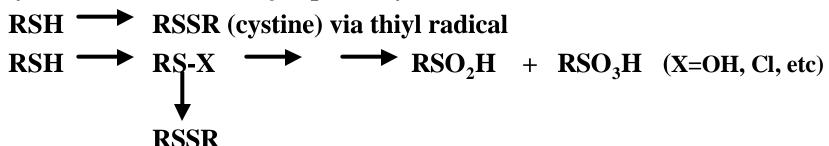
- 1) Decomposition of dimers formed between two peroxyl radicals ROO-OOR  $\longrightarrow$  2RO? + O<sub>2</sub>
- 2) Hydrogen atom abstraction by an initial peroxyl radical ROO? + XH ROOH + X?
- 3) Fragmentation reactions of ? -hydroxperoxy radicals on Ser and Thr
- 4) Decomposition of hydroperoxides to alkoxyl radicals

All of these reactions give rise to further radicals and hence either

- chain reactions on proteins, or
- damage to other biomolecules

## Oxidation of Cys, cystine and Met residues

- Important targets: rapid reaction with range of oxidants (both radical and non-radical) and easily oxidised.
- Met converted to sulfoxide:  $-S- \longrightarrow -S(=O)- \longrightarrow -S(O_2)-$
- Cys oxidised via two major pathways:



- Cystine can be oxidised to RSS(=O)R
- Both Met sulfoxide and cystine can be <u>repaired</u>
- Only major examples of repair of oxidised amino acid residues on proteins

Free Radic Biol Med, 1995, <u>18</u>, 93; PNAS, 2001, <u>98</u>, 12920, Free Radic Biol Med, 1995, <u>31</u>, 1432; Int J Radiat Biol, 1989, <u>55</u>, 539; von Sonntag - The Chemical Basis of Radiation Biology

# Mechanisms of side-chain oxidation by radicals: aromatic residues

Major reaction is addition, though electron abstraction can also occur.

Electron abstraction reactions usually yield hydroxylated products.

Addition reactions tend to yield a greater diversity of products as this depends on the added species.

Similar products tend to be formed in presence and absence of  $O_2$ .

### Specific aromatic side-chain oxidation products

#### Phenylalanine

o-, m-tyrosine

dimers

Phenylalanine

**Tyrosine** 

#### **Tyrosine**

**DOPA** 

di-tyrosine

#### **Tryptophan**

N-formylkynurenine,

kynurenine,

5-hydroxytryptophan,

7-hydroxytryptophan

#### 3,4-dihydroxyphenylalanine (DOPA)

#### Histidine

di-Tyrosine (di-Tyr)

Free Radic. Biol. Med., 1999, <u>27</u>, 1151-1163

## Transfer of damage within proteins

- Evidence for long range transfer of radical sites within proteins:
  - transfer from initial site to a readily oxidised residue (Trp, Tyr, Met,
     Cys) to give more stable radical can be equilibria.
  - can occur over very large distances, but depends on protein structure
  - occurs in competition with reaction of initial radical with  $O_2$
  - most common when initial radical is poorly, or unreactive, with  $O_2$
  - examples:  $Trp \leftarrow Tyr$ ,  $Trp \leftarrow Cys$ ,  $Tyr \leftarrow Cys$

Oxidised porphyrin ring from Trp, Tyr, Cys

J. Am. Chem. Soc., 1989, <u>111</u>, 5141-5145; J. Am. Chem. Soc., 1994, <u>116</u>, 12010-12015; Int. J. Radiat. Biol., 1989, <u>55</u>, 539-556; J. Biol. Chem., 1997, <u>272</u>, 2359-2362

### Mechanisms of protein oxidation by non-radical oxidants

- Species such as <sup>1</sup>O<sub>2</sub>, HOCl, HOBr, peroxynitrite, O<sub>3</sub>, UV light
- Reactions can be very selective and generally on side-chains Little fragmentation, but considerable aggregation
  - ${}^{1}O_{2}$  damage to Cys, Met, Trp, Tyr and His.
  - HOCl / HOBr primarily damage to Cys, Met, His, Lys, Trp,? amino group.
  - Peroxynitrite damage to Cys, Tyr, Trp.
  - UV light damage to cystine, Trp, Tyr and His.
- Some products well-defined e.g. 3-chloroTyr, 3-nitroTyr, methionine sulfoxide, disulphides.
- Some poorly defined -> peroxides generated by  ${}^{1}O_{2}$  and  $O_{3}$ .

# Aromatic side-chain oxidation products generated by non-radical oxidants

Free Radic. Biol. Med., 1999, <u>27</u>, 1151-1163; Photochem. Photobiol, 2002, <u>76</u>, 35-46

# Damage to other targets induced by reactive intermediates of protein oxidation

- Protein peroxides can induce damage to other proteins by:
  - Non-radical reactions (oxidation of Cys and Met residues)
  - Radical-mediated reactions

Biochem. J., 1999, 338, 629-636; Biochem. J., 1999, 344, 125-134; Chem. Res. Toxicol., 2000, 13, 665-672; Biogerentology, 2002, 3, 95-102; Eur. J. Biochem., 2002, 269, 1916-1925; FEBS Lett., 2002, 527, 289-292.

### Oxidation of other residues by non-radical reactions

- Evidence for oxidation of cysteine residues by peroxides
  - Protein thiols, peroxides and enzyme activity lost concurrently
  - Cysteine oxidised to disulphide and oxy acids
- Evidence for oxidation of methionine residues by peroxides
  - Evidence for the generation of methionine sulfoxide
  - Inactivation of methionine-dependent enzymes ?

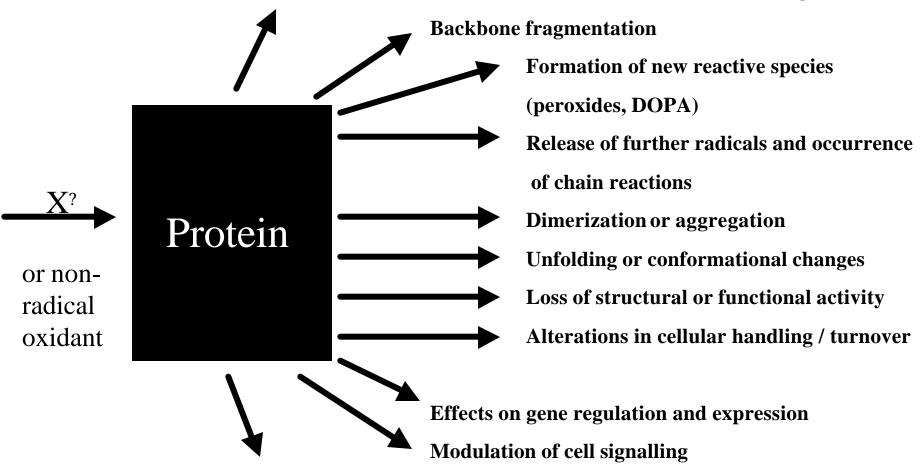
# Damage to other targets induced by reactive intermediates of protein oxidation

- Protein peroxides can induce damage to other proteins by:
  - Non-radical reactions (oxidation of susceptible thiols)
  - Radical-mediated reactions
- Protein peroxides and DOPA can induce damage to DNA via radical-mediated reactions
  - Oxidation of bases (e.g. 8-oxodG)
  - Induction of strand breaks
  - Formation of protein-DNA cross-links
- Protein peroxides can induce lipid oxidation via radicalmediated reactions

Biochem. J., 1999, 338, 629-636; Biochem. J., 1999, 344, 125-134; Chem. Res. Toxicol., 2000, 13, 665-672; Biogerentology, 2002, 3, 95-102; Eur. J. Biochem., 2002, 269, 1916-1925; FEBS Lett., 2002, 527, 289-292.

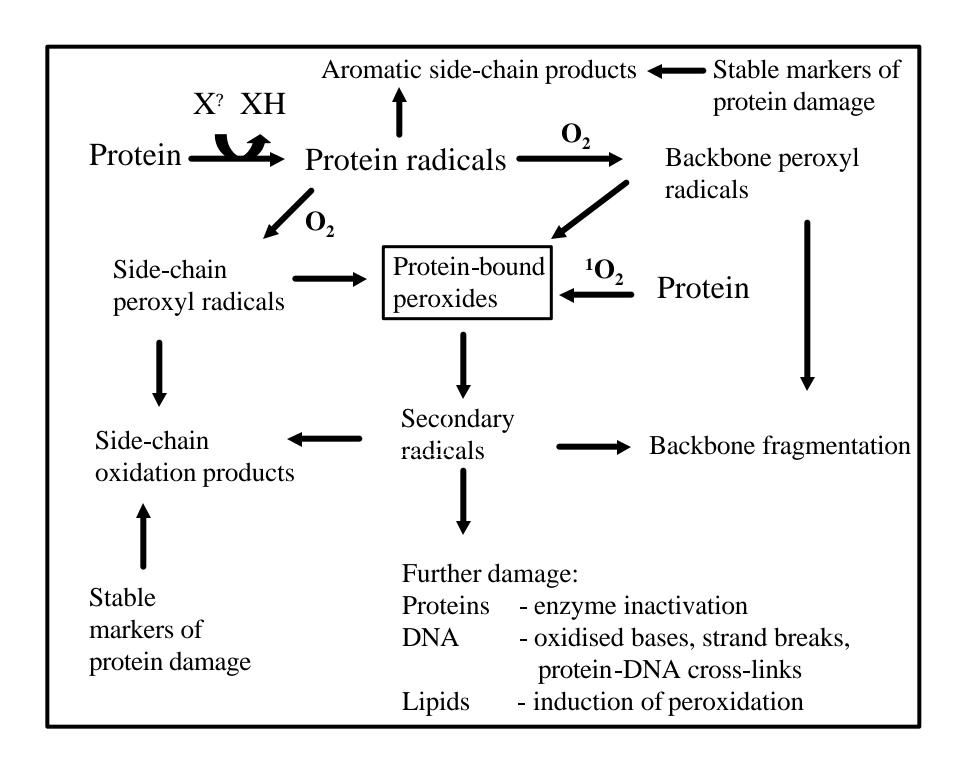
### Consequences of oxidation of proteins

Oxidation of side-chains - some of which can be used as damage markers



Induction of apoptosis and necrosis

Biochim. Biophys. Acta, 2001, <u>1504</u>, 196-219; J. Photochem. Photobiol. B, 2001, <u>63</u>, 114-125.



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